

QUINAZOLINE ALKALOIDS IN NATURE

A. L. D'yakonov and M. V. Telezhenetskaya

UDC 547.944/945

This review generalizes information on the distribution in the plant and animal kingdoms of all types of quinazoline alkaloids known at the present day.

The main structural element of the class of quinazoline alkaloids is the heterocyclic skeleton of quinazoline, which has not hitherto been detected in living Nature and was obtained synthetically in 1895 [1]. The quinazoline alkaloids form a small group of secondary natural compounds. At the present time (up to and including 1995), more than 140 substances of this type are known. The chemistry of the quinazoline alkaloids had its origin in 1888, with the discovery of the first natural representative of them — (+)-peganine (known in the foreign literature as vasicine). The beginning of the 20th century (namely, in 1909 and 1915) saw the isolation of two of the most important parents of other structural types of quinazoline bases — tetrodotoxin and rutecarpine. Then followed the first lull, and only in the mid-1940's and, especially, in the 1950's did a vigorous development of the chemistry of the quinazoline alkaloids begin. It must be mentioned that, in parallel with work on the isolation of this group from various natural sources and the determination of their structures, investigations on their synthesis and the determination of their biological activities were intensively developed, and today the number of synthetic quinazoline derivatives many times exceeds the number of natural quinazoline bases. Correspondingly, the number of publications on this subject is considerable. In view of this, we have limited ourselves to a consideration of the distribution of quinazoline compounds in living Nature.

A number of publications of review nature [2-5] have been devoted to the chemistry of the quinazoline alkaloids; however, in our opinion, they do not give a general picture of the distribution of this class of compounds in the plant and animal kingdoms. Thus, for example, in some publications a particular structural type is considered [6, 7]; in others there is information not only on the natural alkaloids but also on their numerous synthetic analogs [8, 9]; a third group gives the chemical composition of only one plant [10]; and, finally, another group consists of simple lists of facts accumulated over a definite time interval [11-13]. Furthermore, there are no generalizing papers on individual groups of quinazolines. We have attempted to bring together the available information on the quinazoline alkaloids and to show their place in the plant and animal kingdoms.

The distribution in Nature of bases of the quinazoline series is not restricted by the frameworks of any one or more taxonomic units. These alkaloids, unlike many others, are found not only in higher plants (both monocotyledons and dicotyledons) but also in lower plants (fungi and microorganisms) and in the most diverse classes of animals (mainly dwellers of the seas and oceans).

Table 1 gives information on the distribution of quinazoline alkaloids in the plant and animal kingdoms. It must be mentioned that in its compilation we strove to follow strictly the principle of the evolutionary development of living material from simple to complex. All the taxons in which representatives of the quinazoline bases have been detected are arranged in just this sequence. The classification of plant and animal taxons is given in accordance with the schemes adopted in two fundamental monographs [14, 15].

In accordance with the classification of the quinazoline alkaloids by structural types adopted at the present time, we propose to distinguish the following groups within this class of compounds: I) simple (bicyclic) quinazolines; II) tricyclic quinazolines; III) quinazolinocarbolines; IV) tryptoquivalines; V) tetrodotoxins; and VI) quinazolines with various structures. In accordance with this scheme, we shall consider each group individually.

Institute of the Chemistry of Plant Substances, Academy of Sciences of the Republic of Uzbekistan, Tashkent, fax (3712) 40 64 75. Translated from *Khimiya Prirodnikh Soedinenii*, No. 3, pp. 297-351, May-June, 1997. Original article submitted April 1, 1966.

TABLE 1. Distribution of Quinazoline Alkaloids in Nature

Biological species	Literature ref.	Structural type
Lower Plants (bacteria and fungi)		
I. Spirillaceae		
<i>Vibrio alginolyticus</i> and other marine species of the <i>Vibrio</i> genus	113, 121, 123	V
II. Pseudomonadaceae		
<i>Pseudomonas aeruginosa</i> , <i>P. fluorescens</i> , <i>P. putida</i>	11, 13, 14, 17, 18, 19	I
Other marine species of the <i>Pseudomonas</i> genus (marine species)	113, 121, 123	V
<i>Bacillus</i> (marine species)	113, 121, 123	V
<i>Plesiomonas</i> (marine species)	121	V
III. Bacteriaceae		
<i>Klebsiella pneumoniae</i>	43	II
IV. Saccharomycetaceae		
<i>Candida lipolytica</i>	139, 141	VI
V. Deuteromycetes (Imperfect Fungi)		
<i>Alternaria citri</i>	2, 25	I
<i>Cladobotryum varium</i>	2	I
<i>Colletotrichum lagenarium</i>	2	I
VI. Ascomycetes (Ascomycetous Fungi)		
<i>Aspergillus alliaceus</i>	126, 127, 128, 129, 130	VI
<i>A. clavatus</i>	93, 94, 95, 96, 99, 111	IV
<i>A. fischeri</i>	125, 132, 133	VI
<i>A. flavipes</i>	140	VI
<i>A. fumigatus</i>	96, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 143, 144, 145	IV, VI
<i>Corynascus setosus</i>	97, 98, 142	IV, VI
<i>Fusarium culmorum</i>	2	I
<i>Neosartorya fischeri</i>	142	VI
<i>Penicillium aurantiogriseum</i>	131	VI
<i>P. chrysogenum</i>	2, 25	I
<i>Penicillium</i> sp.	134, 135, 136, 137	VI
Higher Plants		
I. Polygonaceae		
<i>Polygonum tinctorium</i>	141	VI
II. Brassicaceae (Cruciferae)		
<i>Isatis tinctoria</i>	139, 141	VI
III. Malvaceae		
<i>Sida acuta</i> , <i>S. cordifolia</i> , <i>S. humilis</i> , <i>S. rhombifolia</i> , <i>S. spinosa</i>	35, 37, 60	II
IV. Saxifragaceae		
<i>Dichroa tebrifuga</i>	12, 21, 22, 23	I
<i>Hydrangea macrophylla</i>	22, 23	I
<i>H. umbellata</i>	12, 22, 23	I
V. Fabaceae (Leguminosae)		
<i>Galega officinalis</i>	37, 46, 48, 61	II
<i>G. orientalis</i>	60	II
VI. Lecythidaceae		
<i>Couropita guianensis</i>	141	VI
VII. Rutaceae		
<i>Analiopsis tabouensis</i>	75, 82	III
<i>Euxylophora paraensis</i>	66, 74, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92	III
<i>Evodia meliaeifolia</i>	70, 76, 82	III
<i>E. rutaecarpa</i>	67, 68, 70, 71, 72, 73, 76, 77, 80, 82	III

TABLE 1 (continued)

Biological species	Literature ref.	Structural type
<i>Glycosmis arborea</i>	1, 5, 6, 7	I
<i>G. pentaphylla</i>	1, 5, 8	I
<i>Hortia arborea</i>	76, 78, 79	III
<i>H. badini</i>	76, 79	III
<i>H. braziliana</i>	78	III
<i>H. longifolia</i>	79	III
<i>H. regia</i>	76	III
<i>Tetradium glabrifolium</i>	66	III
<i>Vepris louisii</i>	66, 69	III
<i>Zanthoxylum arborescens</i>	15, 16	I
<i>Z. budrunga</i>	1, 75, 76	I, III
<i>Z. limonella</i> , <i>Z. pluviatile</i>	76	III
<i>Z. oxyphyllum</i>	75	III
<i>Z. rhetsa</i>	75, 81, 82	III
VIII. Biebersteiniaceae		
<i>B. multifida</i>	37	II
IX. Peganaceae		
<i>Peganum harmala</i>	32, 37, 38, 46, 47, 48, 49, 50, 57, 58, 59, 60, 62	II
<i>P. nigellastrum</i>	37, 46, 48, 60	II
X. Nitrariaceae		
<i>Nitraria komarovii</i>	38, 46, 48, 53, 54, 55, 56	II
<i>N. sibirica</i>	37, 46	II
XI. Araliaceae		
<i>Mackinlaya klossii</i> , <i>M. subulata</i>	64, 65	II
<i>M. macrosciadia</i>	41, 46, 64, 65	II
XII. Rubiaceae		
<i>Galium aparine</i>	37, 38, 42, 44	II
XIII. Scrophulariaceae		
<i>Linaria transiliensis</i>	37, 46, 60	II
<i>L. vulgaris</i>	37, 60	II
<i>L. alpina</i> , <i>L. anticaria</i> , <i>L. avenaria</i> , <i>L. dalmatica</i> , <i>L. genitifolia</i> , <i>L. popovii</i> , <i>L. pseudo-laxiflora</i> , <i>L. purpurea</i> , <i>L. repens</i> , <i>L. triornithophora</i> , <i>L. ventricosa</i> , <i>L. vulgariformis</i> and some other species of the <i>Linaria</i> genus	60	II
XIV. Acanthaceae		
<i>Achatodes beddomei</i>	35, 37, 46, 59	II
<i>A. vasica</i>	27, 28, 29, 32, 33, 34, 35, 36, 37, 39, 40, 45, 48, 51, 59, 60	II
<i>Anisotes sessiliflorus</i>	29, 30, 31, 45, 60, 63	II
<i>Strobilanthes cusia</i>	21, 24, 139, 141	I, VI
XV. Asteraceae (Compositae)		
<i>Echinops echinatus</i>	4, 26	I
XVI. Poaceae (Gramineae)		
<i>Arundo donax</i>	46	II
XVII. Arecaceae (Palmae)		
<i>Daemonorops draco</i>	52	II
Animals		
I. Anthozoa (Corals)		
<i>Anemonia sulcata</i>	121	V
II. Turbellaria		
<i>Planocera multitentaculata</i>	121	V
III. Nemertini		
<i>Lineus fuscoviridis</i>	113, 121, 123	V
<i>Tubulanus punctatus</i>	113, 121	V

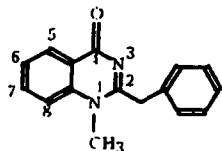
TABLE 1 (continued)

Biological species	Literature ref.	Structural type
IV. Bryozoa		
<i>Hincksinoflustra denticulata</i>	138	VI
V. Gastropoda		
<i>Babylonia japonica</i>	121	V
<i>Charonia sauliae</i>	113, 121, 123	V
<i>Monodonta labio</i>	20	I
<i>Natica lineata</i>	113, 121	V
<i>Niotha clathrata</i>	121	V
<i>Tutufa lissostoma</i>	121	V
<i>Zeuxis siquijorensis</i>	121	V
VI. Cephalopoda		
<i>Octopus maculosus</i>	113, 121, 123	V
VII. Crustacea		
<i>Atergatis floridus</i>	113, 121	V
<i>Zosimus aeneus</i>	121	V
VIII. Merostomata		
<i>Androctonus australis</i>	121	V
<i>Carcinoscorpius rotundicauda</i>	121	V
IX. Myriapoda		
<i>Glomeris marginata</i>	9, 10	I
X. Asteroidea (Starfish)		
<i>Astropecten latespinosus, A. polyacanthus, A. scoparius</i>	121	V
XI. Ascidiacea		
<i>Pyura sarciformis</i>	3	I
XII. Pisces (Fish)		
a) Gobiidae		
<i>Gobius criniger</i>	121	V
b) Tetraodonitidae		
<i>Amblyrhynchotes hypselogenion</i>	121	V
<i>Arothron nigropunctatus</i>	116, 118, 119, 121, 123, 124	V
<i>A. stellatus</i>	121	V
<i>Fugu alboplumbeus, F. oblongus, F. stictionotus</i>	121	V
<i>F. niphobles</i>	116, 118, 121, 124	V
<i>F. pardalis, F. poecilnotus</i>	113, 116, 118, 120, 121, 123, 124	V
<i>F. rubripes</i>	113, 120, 121, 123	V
<i>F. vernicularis</i>	113, 121	V
<i>Lagocephalus lunaris</i>	121	V
<i>Pugilina ternatana</i>	121	V
XIII. Amphibia		
a) Ambystomatidae		
<i>Ambystoma tigrinum</i>	121, 124	V
b) Brachycephalidae		
<i>Atelopus ambulatorius, A. oxyrhynchus, A. senex, A. varius</i>	121	V
<i>A. chiriquiensis</i>	121, 122	V
<i>Brachycephalus ephippium</i>	121	V
c) Salamandridae (True salamanders)		
<i>Cynops ensicauda</i>	112, 113, 114, 116, 117, 121, 123, 124	V
<i>C. marmorata</i>	121	V
<i>C. pyrrhogaster</i>	116, 121, 124	V
<i>C. viridescens</i>	121, 124	V
<i>Paramesotriton hongkongensis</i>	121	V
<i>Taricha granulosa</i>	115, 116, 121	V
<i>T. rivularis, T. torosa</i>	121	V
<i>Triturus alpestris</i>	116, 121, 124	V
<i>T. oregon</i>	116, 121	V
<i>T. vulgaris</i>	121, 124	V

I. Simple (Bicyclic) Quinazolines

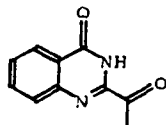
The simple (bicyclic) quinazoline alkaloids are very widely distributed in living Nature. They have been detected in bacteria (*Pseudomonas*), fungi (*Alternaria*, *Cladobotryum*, *Colletotrichum*, *Fusarium*, *Penicillium*), in higher plants of the families Saxifragaceae (*Dichroa*, *Hydrangea*), Rutaceae (*Glycosmis*, *Zanthoxylum*), Asteraceae (*Echinops*), and Acanthaceae (*Strobilanthes*), and in gastropod mollusks (*Monodonta*), Myriopoda (*Glomeris*), and ascidians (*Pyura*). At the present time, 26 representatives of this group of alkaloids are known.

From the chemical point of view these compounds are represented by a quinazoline nucleus, which may contain substituents in positions 1, 2, 3, 4, and 7. These may be methyl, ethyl, hydroxyl, hydroxymethyl, benzyl, carbonyl, carboxyl, amide, bromine, or acetyl. In addition, more complex groupings are often found in position 3. For example, in the ascidian *Pyura sacciformis* an alkaloid containing bromine — 7-bromoquinazoline-2,4-dione — has been detected, which is a fairly rare occurrence.



1. ARBORINE
Glycosmis arborea [16], *G.pentaphylla* [17],
Zanthoxylum budrunga [18]
 $C_{16}H_{14}N_2O$: 250.2990
 mp.: 155–156°
 { picr. 174° (dec.) n-cl. 215°, h.b. 76°, h-i. 96°
 nitr. 117°, picrol. 171°, m-i. 126–127° }

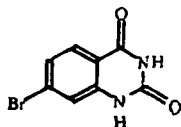
UV: 227, 268, 277, 306, 315 (4.25, 3.68, 3.72, 3.94, 3.87) [188]
 IR: 1645, 1603, 1531, 1502, 1456 [189]
 Mass: 250, 249, 133, 132, 105, 104
 PMR: 3.60 (3H, s, NCH₃); 4.26 (2H, s, PhCH₂); 7.10–7.90 (8H, m, H-Ar); 8.30 (1H, q, J=8, 2, H-5) [190]



2. 2-ACETYLQUINAZOLIN-4(3H)-ONE
Alternaria citri [22], *Cladobotryum varium* [23],
Colletotrichum lagenarium [24], *Fusarium*
culmorum [25], *Penicillium chrysogenum* [26]
 $C_{10}H_{10}N_2O_2$: 188.1846
 mp: 205° (e-a) [24], 200–201° (acetonitrile) [193]

UV: 229, 303 (3.38, 3.13) [23]
 IR: 3165, 3065, 1710, 1600
 Mass: 188 (M^+ , 100), 160 (17), 146 (46), 119 (28) [23]
 PMR: 2.70 (3H, s, CH₃); 7.40–7.90 (3H, m, H-Ar); 8.20 (1H, m, H-Ar); 12.20 (1H, br.s, NH) [193]
¹³C NMR [193]

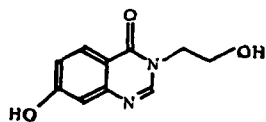
C-9	194.0 (s)	C-7	134.7 (d)	C-5	126.1 (d)
4	159.9 (s)	6	128.8 (d)	4a	123.6 (s)
1a	147.3 (s)	8	128.5 (d)	10	24.7 (q)
2	147.1 (s)				



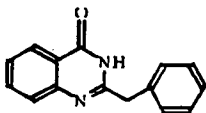
3. 7-BROMOQUINAZOLINE-2,4-DIONE
Pyura sacciformis [33]
 $C_8H_5BrN_2O_2$: 241.0429
 mp.: >330° (meth.)
 UV: 225, 310
 IR: 1730, 1680

Mass: 242 (99), 240 (M^+ , 100), 199 (70), 197 (69), 172 (41), 170 (48)
 PMR: 7.36 (1H, dd, J=2.0, 8.9); 7.39 (1H, d, J=2.0); 7.89 (1H, d, J=8.9)

4. 7-HYDROXYECHINOZOLINONE



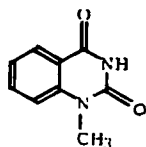
Echinops echinatus [27]
 $C_{10}H_{10}N_2O_3$: 206.1994



Glycosmis arborea [28], G. pentaphylla [29]
 $C_{15}H_{12}N_2O$: 236.2722
 mp.: 249°
 UV: 225, 265, 303, 312 (4.44, 3.95, 3.66, 3.57)
 [188]

IR: 3440, 3356, 1676, 1613, 770, 748, 713
 Mass: 236 (M^+), 235, 119, 92, 91, 90, 78, 77
 PMR: 4.08 (CH_2); 7.50 (8H, H-Ar); 8.26 (d, H-5); 10.25 (NH) [189]

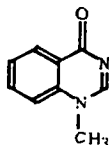
5. GLYCOSMININE



Glycosmis arborea [19]
 $C_9H_8N_2O_2$: 176.1736
 mp.: 270–271° (meth. chl.-meth.)
 UV: 219, 244, 311 (4.70, 3.97, 3.64)
 IR: 1701, 1661, 1605, 1484, 1399 [189]

Mass: 176 (M^+), 133, 132, 105, 104, 92, 90, 78, 77
 PMR: 3.58 (NCH_3); 7.72 (3H, H-6, H-7, H-8); 8.22 (1H, d, H-5); 8.59 (NH)

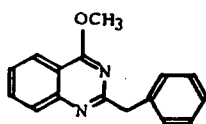
6. GLYCOSMICINE



Glycosmis arborea [19]
 $C_9H_8N_2O$: 160.1746
 mp.: 145–147°
 h-chl. 242° (dec.) picr. 249° (dec.)
 UV: 230, 269, 278, 306, 317 (4.08, 3.59, 3.66, 3.91, 3.84) [188]

UV (H^+) 282, 295, 304 (3.68, 3.73, 3.69) [189]
 IR: 3113, 3067, 3022, 1704, 1652, 1635, 1620, 1540, 1498, 1382, 758
 Mass: 160 (M^+), 133, 132, 105, 104, 92, 90, 78, 77
 PMR ($DMSO-d_6$): 3.83 (NCH_3); 7.35 (H-8); 7.49 (H-6); 7.73 (H-7); 8.25 (H-2), 8.35 (H-5) [189]

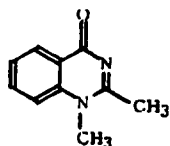
7. GLYCORINE



Glycosmis pentaphylla [35]
 $C_{16}H_{14}N_2O$: 250.2990
 mp.: 165°
 UV: 230, 268, 276, 302, 312 (4.35, 3.85, 3.78, 3.50, 3.45)

IR: 1618, 1610, 1545, 1208
 Mass: 249 (M^+), 235, 119, 91
 PMR: 3.70 (3H, OCH_3); 4.23 (2H, CH_2); 7.25 (5H, H-Ar); 7.50–7.80 (3H, H-Ar); 8.10 (1H, H-5)

8. GLYCOPHYMOLINE

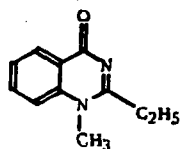


Glomeris marginata [20, 21]
 $C_{10}H_{10}N_2O$: 174.0793
 mp.: 198–200° [20]; 204–205° (e-a.) [21]
 UV: 230, 265, 275, 305, 315
 Mass: 174 (M^+), 146, 145, 133, 105, 104

PMR: 2.63 (3H, s, CH_3); 3.76 (3H, s, NCH_3); 7.20–7.86 (3H, m, H-Ar); 8.23 (1H, dd, J=3, 7, H-5) [191]

9. GLOMERINE

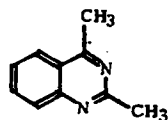
10. HOMOGLOMERINE



Glomeris marginata [20, 21]
 $C_{11}H_{12}N_2O$: 188.0948
 mp.: 145–147° [20]
 149° (e-a) [192]
 UV: 230, 265, 275, 305, 315

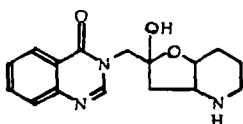
Mass: 188 (M^+), 160, 133, 105
 PMR: 1.41 (3H, t, $J=8$, CH_2CH_3); 2.90 (2H, q, $J=8$, CH_2CH_3); 3.75 (3H, s, NCH_3); 7.33–7.80 (3H, m, H-Ar); 8.33 (1H, dd, $J=3, 7$, H-5) [191]

11. 2,4-DIMETHYLQUINAZOLINE



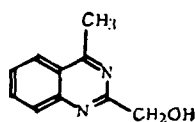
Pseudomonas aeruginosa, *P. fluorescens*, *P. putida*
 [36]
 $C_{10}H_{10}N_2$: 158.2024
 mp.: 63–64°

12. ISOFEBRIFUGINE* [227 a]



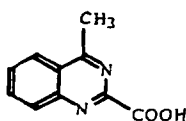
Dichroa febrifuga [37, 38], *Hydrangea umbellata*
 [39]
 mp: 138–139° (ac.)
 $[\alpha]_D^{25} = -121^\circ$ (chl.f.) [226]
 IR: 1695–1678, 1613, 1105, 1055 [226]

13. 2-HYDROXYMETHYL-4-METHYLQUINAZOLINE



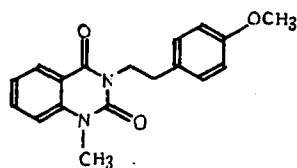
Pseudomonas aeruginosa, *P. fluorescens*, *P. putida*
 [36]
 $C_{10}H_{10}N_2O$: 174.2014

14. 2-CARBOXY-4-METHYLQUINAZOLINE



Pseudomonas aeruginosa, *P. fluorescens*, *P. putida*
 [36]
 $C_{10}H_8N_2O_2$: 188.1846

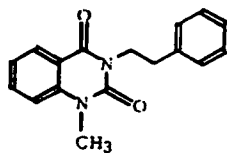
15. 3-(2'-[4"-METHOXYPHENYL)ETHYL]-1-METHYL-1H,3H-QUINAZOLINE-2,4-DIONE



Zanthoxylum arborescens [34]
 $C_{18}H_{18}N_2O_3$: 310.1317
 mp: 133–134° (e-a-pet.eth.)

UV: 222, 242, 276, 283, 310 (61000, -, 3200, 3200, 4100)
 IR: 1700, 1660, 1620
 Mass: 311 (10), 310 (M^+ 45), 155 (11), 135 (100), 134 (52), 121 (75), 119 (54), 58 (20), 57 (16)
 PMR: 2.87 (2H, m, CH_2); 3.58 (3H, s, NCH_3); 3.77 (3H, s, OCH_3); 4.44 (2H, m, CH_2); 6.90, 7.20 (A_2B_2 system); 8.12 (1H, q, H-6)

16. 1-METHYL-3-(2'-PHENYLETHYL)-1H,3H-QUINAZOLINE-2,4-DIONE



Zanthoxylum arborescens [34]
 $C_{17}H_{16}N_2O_2$: 280.1211
 mp.: 100-102° (e-a-pet.eth.)
 UV: 221, 240, 310 (61000, -, 4100)
 IR: 1702, 1655, 1620

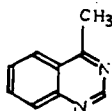
Mass: 280 (M^+ , 67), 189 (28), 177 (35), 176 (100), 134 (27), 133 (28), 132 (15), 105 (40), 104 (77), 91 (15), 78 (17), 77 (27), 56 (15)

PMR: 2.93 (2H, m); 3.55 (3H, s, NCH_3); 4.40 (2H, m, CH_2); 7.80 (J=7, H-Ar); 8.12 (1H, q, H-6)

^{13}C NMR:

C-2	140.4	C-8	134.9	C-13	138.6
4	161.4	9	113.3	16	126.4
5	115.4	10	150.7	17	128.4*
6	128.9	11	34.0	18	128.9*
7	122.8	12	43.2	NCH_3	30.6

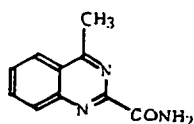
17. 4-METHYLQUINAZOLINE



Pseudomonas aeruginosa, P.fluorescens, P.putida
 [36]

$C_9H_8N_2$: 144.1756
 mp.: 36-37°

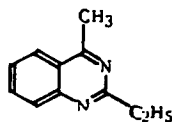
18. 4-METHYLQUINAZOLINE-2-CARBOXAMIDE



Pseudomonas aeruginosa, P.fluorescens, P.putida
 [36]

$C_{10}H_9N_3O$: 187.2002

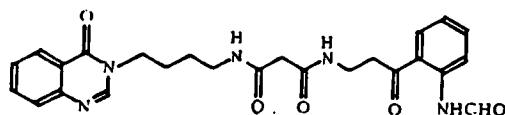
19. 2-ETHYL-4-METHYLQUINAZOLINE



Pseudomonas aeruginosa, P.fluorescens, P.putida
 [36]

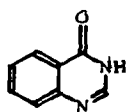
$C_{11}H_{12}N_2$: 172.2292

20. MONODONTAMIDE F



Monodonta labio [40]
 $C_{25}H_{27}N_5O_5$: 477.5168

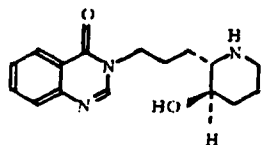
21. 4-OXOQUINAZOLINE



Dichroa febrifuga [30], Strobilanthes cusia [194]
 $C_8H_6N_2O$: 146.1478
 mp: 212–213° (alc.)
 h-chl. 247° (eth.)

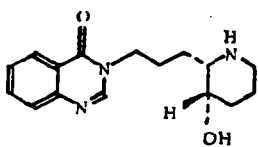
UV: 223, 265, 300, 313 (4.36, 3.81, 3.60, 3.54) [188]
 IR: 1664, 1608 [197]
 Mass: 146 (M^+) [221]

22. *trans*-FEBRIFUGINE



Hydrangea macrophylla [195]
 $C_{16}H_{21}N_3O_2$: 287.3600

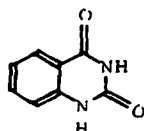
23. *cis*-FEBRIFUGINE* [227 b]



Dichroa febrifuga [37, 38], Hydrangea macrophylla [195], H.umbellata [39]
 $C_{16}H_{21}N_3O_2$: 287.3600
 mp: 158–160° (chlif.-pet. eth.)
 $[\alpha]_D^{+16}$ (alc.)

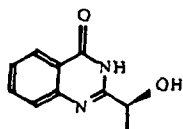
h-chl. 232–233° (meth.-HCl, dec.)
 $[\alpha]_D^{-12.8}$ (water) [226]
 IR: 1727, 1695–1678, 1613 [226]

24. QUINAZOLINE-2,4-DIONE



Strobilanthes cusia [194]
 $C_8H_6N_2O_2$: 162.1468
 mp.: 350°

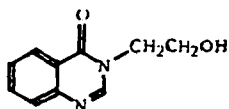
25. CHRYSOGINE



Alternaria citri [22], Penicillium chrysogenum [31]
 $C_{10}H_{10}N_2O_2$: 190.2004
 mp.: 190–192° (e-a)
 $[\alpha]_D^{-41}$ (alc.) [193]; -24 ° [31]
 UV: 225, 230, 265, 271, 305, 316 (27500, 25500, 8250, 7500, 4200, 3450)

IR: 3370, 3120, 2875, 1675, 1625, 1605, 1050 [222]
 PMR: [31]

26. ECHINOSOLINONE



Echinops echinatus [32]
 $C_{10}H_{10}N_2O_2$: 190.2004
 mp.: 150° (chlif.-meth.)
 UV: 230, 265, 270, 319 (4.39, 3.67, 3.68, 3.77)
 IR: 3320, 2920, 2880, 1665, 1620, 1572, 1555, 1495, 1212, 1050, 925, 840

Mass: 190 (M^+), 172, 146, 119, 91
 PMR(CD_3OD): 3.20 (CH_2OH); 4.12 (2H, NCH_2); 7.40–7.65 (m, H-6, H-7, H-8); 8.00 (1H, H-5); 8.80 (H-2)

II. Tricyclic Quinazolines

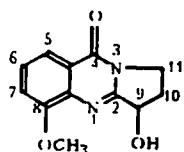
The tricyclic quinazoline alkaloids are found only in higher plants of the following families: Malvaceae (*Sida*), Fabaceae (*Galega*), Biebersteiniaceae (*Biebersteinia*), Peganaceae (*Peganum*), Nitrariaceae (*Nitraria*), Araliaceae (*Mackinlaya*), Rubiaceae (*Galium*), Scrophulariaceae (*Linaria*), Acanthaceae (*Adhatoda*, *Anisotes*), Poaceae (*Arundo*), and Arecaceae (*Daemonorops*). An exception is formed by 8-hydroxydeoxyvasicinone, isolated from a culture of the bacterium *Klebsiella pneumoniae* var. *oxytoca* (*Bacteriaceae*). The tricyclic quinazolines are very numerous. At the present day, 39 compounds of this type are known. The richest sources of them are *Adhatoda vasica* (16 bases isolated), *Peganum harmala* (13 bases), and *Nitraria komarovii* (8 bases).

From the chemical point of view, this group is extremely inhomogeneous. Here it is possible to distinguish "classical" tricyclic quinazoline alkaloids having substituents in position 4 (keto group, hydroxyl, acetone residue), 6 (hydroxyl), 8 (methoxyl, hydroxyl), 9 (hydroxyl), and 11 (hydroxyl). These compounds are mainly characteristic for plants of the genera *Adhatoda*, *Sida*, *Peganum*, *Galium*, *Galega*, and *Linaria*. There are a number of alkaloids containing in position 9 a residue of methyl or ethyl anthranilate or N-methylantranilate and also of N-dimethylaniline (alkaloids of the *Adhatoda* and *Anisotes* genera). There is one representative in each case with the cleavage of ring B (vasicol) and ring C (pegamine). N-oxides of tricyclic quinazoline bases have been isolated from *Nitraria komarovii* quite recently [78-80]. This is the first case of the reliable detection of compounds with groupings of this type among the quinazoline alkaloids. A single indirect mention of this is contained in [88], which also reports the finding of peganine and vasicinone glycosides in an aqueous extract from *Adhatoda vasica*. However, these glycosides were not isolated in the individual state and their structures remain unsubstantiated, for which reason we do not give them.

Quinazolines with a six-membered ring C and cleavage of the bond between rings B and C have been isolated from plants of the Australian genus *Mackinlaya*. And, finally, dimers (dipepine and nordine) have been detected among compounds of the group under consideration.

Some alkaloids of this group are narrowly specialized products of secondary metabolism (for example, the above-mentioned *Anisotes* and *Mackinlaya* alkaloids), while others, conversely, are widely distributed in the vegetable kingdom. For example, (\pm)-vasicinone is found in plants of 8 genera and 15 species, deoxyvasicinone in 7 genera and 9 species, and (\pm)-peganine in 6 genera and 24 species. Furthermore, in a number of cases, tricyclic quinazolines have been isolated from families that, according to some workers [89, 90], cannot in any way be called alkaloid-bearing (Malvaceae, Aralaceae, Arecaceae).

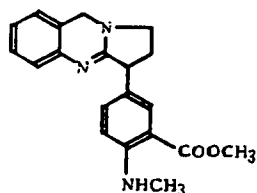
27. ADHAVACINONE



Adhatoda vasica [41]
 $C_{12}H_{12}N_2O_3$: 232.2372
 mp: 230-232°
 $[\alpha]_D^{20}$
 UV: (meth): 214, 238, 284, 314, 326 (4.60, 4.01, 4.21, 3.80, 3.62)

IR: 3260, 1670, 1627, 1570, 1480, 1260, 850
 Mass: 232 (M^+), 217 ($M-15$)⁺
 PMR: 2.35 (2H, m, H-2); 3.88 (3H, s, OCH_3); 4.25 (2H, m, H-1); 5.20 (1H, t, J=7, H-3); 7.20-7.40 (2H, m, H-6, H-7); 7.90 (1H, dd, J=8, 2, H-8)

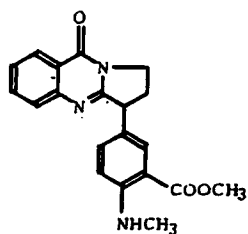
28. ADHATODINE



Adhatoda vasica [83]
 $C_{20}H_{21}N_3O_2$: 335.4040
 mp: 183°
 UV: 225, 262, 300, 361 (4.51, 4.03, 3.86, 3.70)
 IR (chlf.): 3378, 1678, 1626, 1608, 1575, 1439
 IR: 3385, 1685, 1630, 1598, 1575, 770

Mass: 335 (M^+ , 100), 334 (78), 302 (11), 277 (14), 276 (14), 248 (10), 247 (11), 246 (12), 218 (9), 171 (28), 150.5 (21), 136.5 (14)

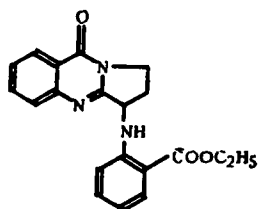
29. ANOSITINE



Adhatoda vasica [83], Anisotes sessiliflorus [69]
 $C_{20}H_{19}N_3O_3$: 349.3872
 mp.: 189–190° (ac.-hx.) [69]; 186° [83]
 (3-hydroxy 185–187° (meth.)
 UV: 212, 226, 259, 301, 312, 356 (37700, 54100,
 14800, 4900, 4500, 5400)
 IR: 2.99 μ , 6.09 μ , 5.95 μ
 Mass: 349 (M^+), 348, 317, 316, 290, 262, 185,
 158.5

PMR (τ -scale): 1.73 (bd, J=9, H-8); 2.17 (d, J=3, H-2'); 3.36 (d, J=9, H-5'); 5.40–6.10 (3H, H-3, H-5); 6.20 (s, OCH₃); 7.12 (s, NCH₃); 7.20–8.00 (m, H-4) [69]

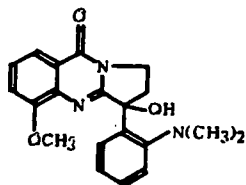
30. ANISESSINE



Anisotes sessiliflorus [69]
 $C_{20}H_{19}N_3O_3$: 349.3872
 mp.: 170–171° (meth.)
 UV: 207, 225, 253, 300, 311, 340 (36600, 54100,
 14700, 4900, 4200, 6300)
 IR: 2.95 μ , 6.05 μ , 5.95 μ
 Mass: 349 (M^+), 303, 302, 275, 200, 183

PMR (τ -scale): 1.73 (bd J=4, H-8); 2.00 (dd, J=4.1, H-3'); 3.10 (d, J=4, H-6'); 3.27 (t, J=4, H-4'); 4.97 (1H, m, H-3); 5.30–6.20 (m, H-5); 7.10–7.84 (dm H-4)

31. ANIFLORINE

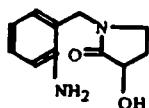


Anisotes sessiliflorus [69]
 $C_{20}H_{21}N_3O_3$: 351.1403
 mp.: 195–197° (meth.)
 UV: 207, 235, 286, 312, 324 (31300, 19900, 7000,
 6600, 5800)
 UV (H⁺): 207, 236, 291, 312, 324 (30500, 16100,
 6200, 7700, 7500)
 IR: 2.90 μ , 6.02 μ

Mass: 351 (M^+), 318, 303, 231, 191, 161, 132

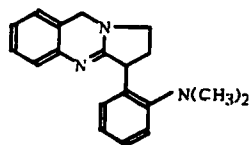
PMR (τ -scale): 2.05 (dd, J=8.1, H-8); 2.70–2.95 (H-3', H-4', H-5', H-6'); 5.40–5.90 (m, H-5); 6.12 (s, OCH₃); 7.20 (s, NCH₃); 7.00–7.50 (m, H-4)

32. VASICOL* [227 c]



Adhatoda vasica [81], Peganum harmala [82]

33. VASICOLINE



Adhatoda vasica [83]
 $C_{19}H_{21}N_3$: 291.1395
 mp.: 135° (ac.-heptane)
 UV: 212, 224, 293 (4.10, 4.07, 3.81)
 IR (chl.f.): 1621, 1592, 1572
 IR: 1652, 1590, 1570, 770, 720

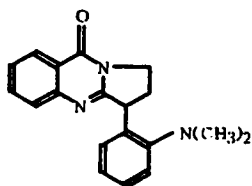
Mass: 291 (M^+), 276, 259, 247, 185, 171, 169, 144, 130

PMR: 1.60–2.10 (2H, m, H-4); 2.67 [s, N(CH₃)₂]; 2.25–3.80 (2H, m, H-4); 3.10–3.40 (2H, m, H-5); 4.50 (1H, t, J=9, H-3); 4.60 (2H, s, H-7); 6.80–7.30 (8H, m, H-Ar)

¹³C NMR [198]

164.9	127.6	50.0
153.1	124.5	47.6
143.6	123.7	46.0 (2C)
137.6	120.8	43.8
128.9	119.3	29.8
128.2		

34. VASICOLINONE



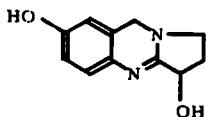
Adhatoda vasica [83]
C₁₉H₁₉N₃O: 305.3782
mp.: 152° (hx.) [83]; 155–156° [196]
UV: 227, 268, 305, 317 (4.33, 3.91, 3.52, 3.42)
IR (chlf.): 1664, 1618 [83]; 1665, 1615 [196]
IR: 1705, 1670, 1620, 785, 765
Mass: 305 (M⁺), 290, 273, 261, 185, 144, 77

PMR: 1.90–2.90 (2H, m, H-4); 4.00–4.55 (2H, m, H-5); 5.04 (1H, t, J=9, H-3); 7.05–7.70 (7H, m, H-Ar): 8.31 (1H, bd, J=9, H-8) [83]

¹³C NMR [198]

161.7.	129.1	121.6
161.2	128.4	120.4
153.2	127.2	46.0 (2C)
149.5	126.2	45.6
136.7	126.0	45.0
133.8	125.0	29.7

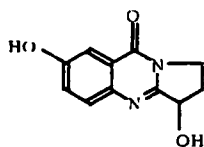
35. VASICINOL



Adhatoda beddomei [42], A. vasica [43], Sida acuta,
S. humilis, S. rhombifolia, S. spinosa [44],
S. cordifolia [45]
C₁₁H₁₂N₂O₂: 204.2272
mp.: 272–273°

[α]_D+46° (c-a)
{6-OMe. 210°}

36. VASICINOLONE

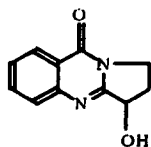


Adhatoda vasica [46]
C₁₁H₁₀N₂O₃: 218.2104
mp.: 279°
{di-Ac. 204°}
UV(meth) 225, 276, 325
IR: 3400, 1670, 1624, 1600, 1490

Mass: 218 (M⁺, 100), 217 (7.7), 190 (5.9), 162 (83.3), 135 (53.1), 131 (13.4), 119 (16.4), 106 (14.2), 69 (57.5)

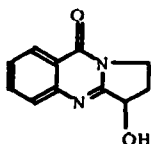
PMR (CD₃COOD): 2.40 (2H, m, H-2); 4.20 (2H, m, H-1); 5.60 (1H, t, H-3); 7.60 (3H, m, H-5, H-6, H-8)

37. (-)-VASICINONE* [227 d]



Adhatoda beddomei [42], A. vasica [47],
Biebersteinia multifida [48], Galega officinalis
[49], Galium aparine [50], Linaria transiliensis
[51], L. vulgaris [52], Nitraria sibirica [53],
Peganum harmala [54], P. nigellastrum [55],
Sida acuta, S. humilis, S. rhombifolia,
S. spinosa [44], S. cordifolia [45]

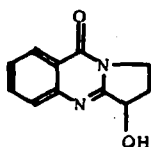
38. (±)-VASICINONE* [227 e]



Galium aparine [50], *Nitraria komarovii* [56],
Peganum harmala [57]

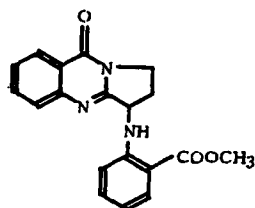
PMR: 2.26 (1H, J=13.4, 8.4, 7.2, 6.8, H-10e); 2.63 (1H, J=13.4, 7.6, 7.3, 4.7, H-10a); 4.03 (1H, J=12.3, 8.4, 4.7, H-11e); 4.33 (1H, J=12.3, 7.6, 7.2, H-11a); 5.16 (1H, J=7.3, 6.8, H-9); 7.51 (1H, J=8.1, 6.8, 1.3, H-6); 7.71 (1H, J=8.2, 1.3, 1.2, H-8); 7.78 (1H, J=8.2, 6.8, 1.3, H-7); 8.28 (1H, J=8.1, 1.3, 1.2, H-5) [117]

39. (+)-VASICINONE



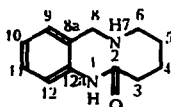
Adhatoda vasica [58]
C₁₁H₁₀N₂O₂: 202.2114
mp.: 201–202° (chlf.-e-a)
[α]_D+148° (alc.)
{Ac. 135°, [α]_D+10° (chlf.)}

40. VASNETINE



Adhatoda vasica [84]
C₁₉H₁₇N₃O₃: 335.3604

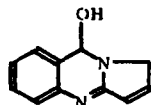
41. 3,4,5,6,7,8-HEXAHYDRO-1,7-BENZODIAZEPIN-2(1H)-ONE



Mackinlaya macrosciadia [86]
C₁₂H₁₆N₂O: 204.2708
mp.: 92.5–93.5° (CCl₄)
Mass: 204 (M⁺)

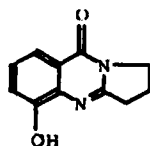
PMR: 1.65–1.85 (4H, m, H-4, H-5); 2.35–2.45 (2H, m, H-3); 3.17–3.28 (2H, m, H-6); 4.65 (2H, m, H-1, H-7); 4.50 (2H, s, H-8); 6.58–7.15 (4H, m, H-9, H-10, H-11, H-12)

42. (–)-8-HYDROXY-2,3-DEHYDRODEOXYPEGANINE



Galium aparine [50]
C₁₁H₁₀N₂O: 186.2124
mp: amorph.
[α]_D–44°

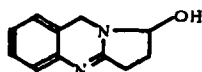
43. 8-HYDROXYDEOXYVASICINONE



Klebsiella pneumoniae [187]
C₁₁H₁₀N₂O₂: 202.2035
mp.: 157° [187]; 158–160° [199]
UV (meth.) 205, 235, 242, 284, 315, 327 (22100,
22100, 18900, 5880, 5830, 5020)

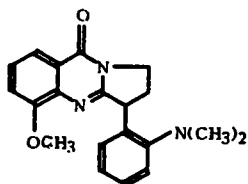
IR (NaCl): 1660, 1620
 Mass: 202.0735 (M⁺, 100), 201 (45), 174.0792 (C₁₀H₁₀N₂O, 11), 145.0764 (C₉H₉N₂, 24),
 145.0524 (C₉H₇NO), 145.0404 (C₈H₅N₂O), 106.0289 (C₆H₆NO, 17), 69.0578 (C₄H₇N, 18)
 PMR (meth. chl.): XCH₂CH₂CH₂Y (2.23, quintet; 3.12, τ; 4.13, t; J=7.4, 8.2); arom. ABX
 system (H_a 7.22, H_b 7.35, H_x 7.64, J_{AB}=7.83, J_{BX}=8.49, J_{AX}=1.83)
¹³CNMR:

160.5	126.7	46.5
158.5	120.4	32.1
151.1	116.9	19.4
137.5	116.6	



44. (-)-1-HYDROXYDEOXYPEGANINE

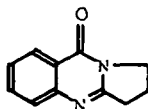
Galium aparine [50]
 C₁₁H₁₂N₂O: 188.2282



45. DEOXYANIFLORINE

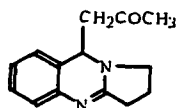
Adhatoda vasica [83], Anisotes sessiliflorus [69]
 C₂₀H₂₁N₃O₂: 335.4040
 mp.: 168–172° (meth.)
 UV: 211, 232, 285, 315, 330 (33000, 23100, 8600,
 6900, 5800)
 UV (H⁺): 211, 237, 285, 315, 320(33000, 20300,
 7700, 6900, 5700)
 Mass: 335 (M⁺), 320, 304, 215, 200, 144

PMR (τ-scale) : 2.12 (dd, J=8.1, H-8), 2.70–3.10 (H-3', H-4', H-5', H-6'): 5.05 (dd, J=10.6, H-3); 5.60–5.95 (m H-5); 6.15 (s, OCH₃); 7.41 (s, NCH₃); 7.10–8.15 (m, H-4)



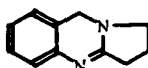
46. DEOXYVASICINONE* [227 f]

Adhatoda beddomei [42], Arundo donax [59], Galega officinalis [52], Linaria transiliensis [51], Mackinlaya macrosciadia [60], Nitraria komarovii [56], N. sibirica [53], Peganum harmala [54], P. nigellastrum [55]



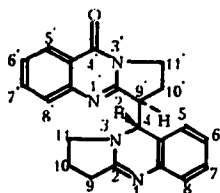
47. DEOXYPEGANIDINE* [227 g]

Peganum harmala [61]



48. DEOXYPEGANINE* [227 h]

Adhatoda vasica [62], Galega officinalis [52], Nitraria komarovii [56], Peganum harmala [62], P. nigellastrum [63]

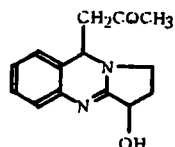


49. DIPEGINE* [227 i]

Peganum harmala [64]

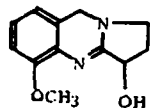
PMR: 1.91 (2H, H-10); 2.10 (1H, J=13.4, 9.0, 8.9, 8.2, H-10'a); 2.20 (1H, J=13.4, 8.9, 8.2, 4.3, H-10'e); 2.61 (1H, J=16.5, 6.5, 6.5, H-9e); 2.70 (1H, J=16.5, 9.5, 9.5, H-9a); 2.89 (1H, J=10.1, 7.8, 7.8, H-11a); 3.22 (1H, J=10.1, 5.9, 5.9, H-11e); 3.55 (1H, J=8.9, 8.9, 2.1, H-9'); 3.96 (1H, J=12.4, 8.2, 8.2, H-11'a); 4.19 (1H, J=12.4, 9.0, 4.3, H-11'e); 5.74 (1H, J=2.1, H-4); 7.08 (1H, J=8.0, 1.5, H-5); 7.10 (1H, J=7.9, 6.9, 1.5, H-7); 7.16 (1H, J=7.9, 1.8, H-8); 7.24 (1H, J=8.0, 6.9, 1.8, H-6); 7.51 (1H, J=8.0, 6.7, 1.6, H-6'); 7.74 (1H, J=8.2, 1.6, 0.6, H-8'); 7.80 (1H, J=8.2, 6.7, 1.4, H-7'); 8.30 (1H, J=8.0, 1.4, 0.6, H-5') [117]
 Abs. conf. 4R, 9'S [117]

50. ISOPEGANIDINE* [227 j]



Peganum harmala [64]

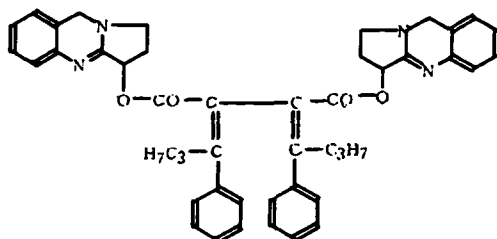
51. 8-METHOXYPEGANINE



Adhatoda vasica [65]
 $C_{12}H_{14}N_2O_2$: 218.2540
 mp.: 224–225°
 $[\alpha]_D^{20}$
 UV: 307 (3.85)

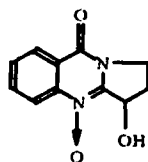
IR: 3470, 1630, 1605, 1500, 1250, 845
 Mass: 218 (M^+), 217, 203, 199
 PMR: 2.18 (2H, m, H-10); 3.21 (2H, m, H-11); 3.80 (3H, s, OCH_3); 4.50 (2H, s, H-4); 4.71 (1H, t, H-9); 6.60–7.00 (3H, m, H-5, H-6, H-7)

52. NORDINE



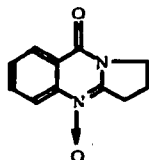
Daemónorops draco [87]
 $C_{46}H_{46}N_4O_4$: 718.8922
 mp.: 292°

53. VASICINONE N-OXIDE* [227 k]



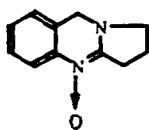
Nitraria komarovii [78]

54. DEOXYVASICINONE N-OXIDE* [227 l]

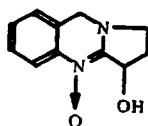


Nitraria komarovii [78]

55. DEOXYPEGANINE N-OXIDE* [227 m]



Nitraria komarovii [79]

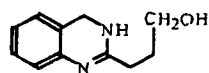


56. PEGANINE N-OXIDE

Nitraria komarovii [80]
 $C_{11}H_{12}N_2O_2$: 204.2272
 mp.: 207–208° (alc.-ac.)
 $[\alpha]_D^{20}$

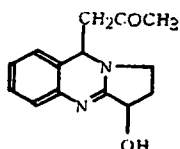
UV: 207, 220 sh., 225, 232 sh., 302 (4.16, 4.21, 4.27, 4.18, 4.06)
 UV (H⁺): 213, 220, 225 sh., 282
 IR: 3150, 2940, 2860, 2840, 1630, 1585, 1510, 1465, 1270, 1230, 770.
 Mass: 204 (M⁺, 3), 203 (4), 188 (8), 187 (12), 186 (10), 171 (9), 155 (26), 140 (100), 123 (24),
 122 (14), 98 (64), 84 (65), 83 (77).
 PMR: 2.25, 2.66 (2H, m, H-10); 3.68 (2H, m, H-11); 4.75 (2H, s, H-4); 5.38 (1H, t, H-9); 6.15
 (1H, br.sOH); 6.96 (1H, m, H-5); 7.16 (3H, m, H-6, H-7, H-8)

57. PEGAMINE* [227 n]



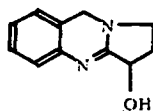
Peganum harmala [75]

58. PEGANIDINE* [227 o]



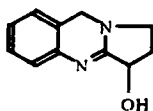
Peganum harmala [62]

59. (-)-PEGANINE [227 p]



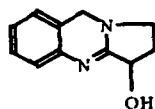
Adhatoda beddomei [42], *A. vasica* [66], *Nitraria komarovii* [56], *Peganum harmala* [67]

60. (±)-PEGANINE* [227 r]



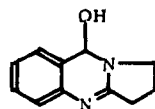
Adhatoda vasica [68], *Anisotes sessiliflorus* [69], *Galega orientalis* [70], *Linaria alpina*, *L. anticaria*, *L. avenaria*, *L. pseudolaxiflora*, *L. repens*, *L. triornithophora* [71], *L. dalmatica*, *L. genistifolia* [72], *L. purpurea*, *L. ventricosa* [73], *L. popovii*, *L. transiliensis*, *L. vulgariformis* [51], *L. vulgaris* [74], *Peganum harmala* [75], *P. nigellastrum* [55], *Sida acuta*, *S. humilis*, *S. rhombifolia*, *S. spinosa* [44], *S. cordifolia* [45]

61. (+)-PEGANINE



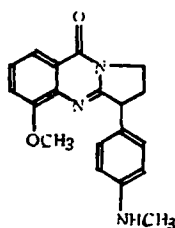
Galega officinalis [76].
 $C_{11}H_{12}N_2O$: 188.2282
 mp.: 170–173°
 $[\alpha]_D^{25} +163^\circ$

62. PEGANOL* [227 s]



Peganum harmala [77]

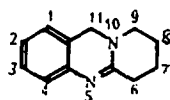
63. SESSIFLORINE



Ansotes sessiflorus [69]
 $C_{19}H_{19}N_3O_2$: 321.3772
 mp.: 195–197° (meth.)
 UV: 212, 239, 286, 315, 327 (34700, 30300, 9200,
 8800, 6600)
 UV (H⁺): 212, 239, 286, 315, 327 (30100, 19400,
 7000, 6900, 5900)
 IR: 6.05 μ
 Mass: 321 (M⁺), 306, 303, 277, 215, 191, 144

PMR (τ -scale): 2.11 (dd, J=8.1, H-8); 2.40–2.80 (m, H-3', H-5'); 3.21 (bd, J=6, H-2'); 3.21 (bd, J=6, H-8'); 5.20–5.85 (3H, m, H-3, H-5); 6.06 (s, OCH₃); 7.05 (s, NCH₃); 7.10–7.60 (m, H-4)

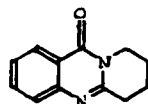
64. 6,7,8,9-TETRAHYDRO-11H-PYRIDO-[2,1-b]-QUINAZOLINE



Mackinlaya klossii [60], *M. macrosciadia*,
M. subulata [85]
 $C_{12}H_{14}N_2$: 186.2560

mp.: 85–87° (eth.)
 [sulf.]: 285° (alc.) picr.: 217°
 UV: 242, 294 (3.78, 3.36)
 IR (CCl₄): 3500, 3380, 3270, 1627 [60]
 Mass: 186 (85), 185 (100), 144 (13), 89 (13), 77 (25), 51 (29)
 PMR: 1.65–1.93 (4H, H-7, H-8); 2.28–2.73 (2H, m, H-6); 2.80–3.17 (2H, m, H-9); 4.33 (2H, s, H-11); 6.67–7.13 (4H, m, H-1, H-2, H-3, H-4) [200]

65. 6,7,8,9-TETRAHYDROPYRIDO-[2,1-b]-QUINAZOLIN-11-ONE



Mackinlaya klossii [60], *M. macrosciadia*, *M. subulata*
 [85]
 $C_{12}H_{12}N_2O$: 200.2392
 mp.: 98.5–99.5° (hx.)

Mass: 200 (M⁺, 100), 199 (85), 185 (54), 130 (13), 116 (21), 90 (27), 89 (21), 77 (25), 76 (28) [200]
 PMR: 2.00 (t, H-7, H-8); 2.80–3.15 (2H, t, H-6); 3.90–4.20 (2H, t, H-9); 7.42–7.63 (H-2, H-3, H-4); 8.12–8.33 (1H, m, J=8.2, H-1)

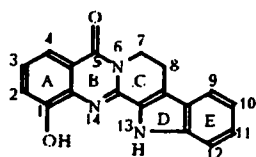
III. Quinazolinocarbolines

The quinazolinocarboline alkaloids form a very specific group of compounds found only in tropical representatives of the family Rutaceae (the genera *Araliopsis*, *Euxylophora*, *Evodia*, *Hortia*, *Tetradium*, *Vepris*, and *Zanthoxylum*). At the present time they number 27 compounds. The richest sources of them are *Euxylophora paraensis* (12 alkaloids) and *Evodia rutaecarpa* (10 alkaloids). As is known, in plants of the Rutaceae family the quinazolinocarbolines are present together with other classes of alkaloids — in particular, those of the furanoquinoline, acridone, and quinoline types. They all have a common anthranilic acid mechanism of biosynthesis. A special investigation has been devoted to questions of the distribution of alkaloid-bearing plants in this family [91]. Within this group, individual genera and species are characterized by the types of alkaloids that they contain. This is shown particularly clearly for species of the genera *Fagara* and *Zanthoxylum*, where the presence of certain bases is not a bad auxiliary taxonomic index. In light of these results and also of information on the distribution of quinazolinocarboline alkaloids over the genera in the Rutaceae family, the possibilities of the chemotaxonomy of this family (particularly the *Evodia* and *Euxylophora* genera) appear to us to be extremely promising.

The quinazolinocarbolines represent a chemical combination of two types of heterocyclic systems: indole and quinazoline. Among them are found representatives with cleavage of ring *B* or *C* and also with the cleavage of rings *B* and *C* simultaneously.

In a number of cases, moreover, ring *C* is dehydrogenated at the C_7-C_8 bond. Substituents may be present in rings *A* (hydroxyl, methoxyl, methylenedioxy) and *E* (methoxyl). The alkaloid paraensine is distinguished by an original structure, which includes an additional partially dehydrogenated pyran ring.

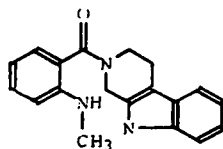
66. 1-HYDROXYRUTECARPINE



Euxylophora paraensis [92], *Tetradium grabrifolium* [94], *Vepris louisii* [93]
 $C_{18}H_{19}N_3O_2$: 303.3188
 mp.: 316–318° (meth.)
 h-chl. 326–328° (meth.-chlif.)

UV (meth+NaOH): 309, 393 [92]
 UV: 286, 293, 332, 348, 361, 279 (3.78, 3.83, 4.34, 4.45, 4.53, 4.42)
 IR: 3340, 3275, 1660, 1630, 1607, 1570, 1533, 1500, 1440, 1394, 1322, 720 [93]
 Mass: 303 (M^+ , 100), 302 (72), 169 (3.8), 168 (9.4)
 PMR: 3.31 (2H, t, $J=7$, H-8); 4.57 (2H, t, $J=7$, H-7); 7.12 (1H, dt, $J=8.1$, H-11); 7.21 (1H, dd, $J=8.1$, H-2); 7.32 (1H, t, $J=8$, H-3); 7.33 (1H, dt, $J=8.1$, H-10); 7.53 (1H, dd, $J=8.1$, H-12); 7.66 (1H, dd, $J=8.1$, H-4); 7.72 (1H, dd, $J=8.1$, H-9); 8.51 (1H, s, OH) [94]

67. GOSUIAMIDE I

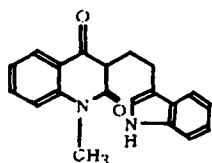


Evodia rutaecarpa [115]
 $C_{19}H_{18}N_2O$: 304.3703
 mp.: 178–180° (bz.-ac.)
 UV: 225, 253, 281.5, 290, 313 (4.65, 4.04, 3.90, 3.84, 3.47)
 IR: 3400, 3280, 2900, 1620, 1500, 1350, 1300, 1020, 990, 890, 740

Mass: 305 (M^+)
 PMR (Py- d_5): 2.69 (3H, d, $J=5.1$, $NHCH_3$); 2.84 (2H, m, H-6); 3.85 (2H, m, H-6); 4.99–5.08 (2H, m, H-3); 6.08 (1H, q, $J=5.1$, H-14); 6.75 (1H, d, $J=8.1$, H-16); 6.80 (1H, t, $J=7.3$, H-18); 7.25–7.33 (2H, m, H-10, H-11); 7.36–7.40 (1H, m, H-17); 7.37 (1H, d, $J=7.3$, H-19); 7.57 (1H, d, $J=8.1$, H-12); 7.56 (1H, d, $J=7.3$, H-9)

^{13}C NMR (Py- d_5):

C-2	131.4 (s)	C-10	119.4 (d)	C-17	131.3 (d)
3	41.8 (t)	11	121.6 (d)	18	115.8 (d)
5	42.3 (t)	12	111.7 (d)	19	128.1 (d)
6	22.1 (t)	13	137.4 (s)	20	120.7 (s)
7	108.0 (s)	15	148.2 (s)	21	171.0 (s)
8	127.7 (s)	16	111.1 (d)	$NHCH_3$	30.0 (q)
9	118.3 (d)				



Evodia rutaecarpa [115]

C₁₉H₁₇N₃O₂: 319.3614

mp.: 216–218° (alc.)

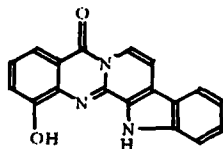
IR 3330, 1690, 1650, 1610, 1490, 1430, 1400,
1360, 860, 840Mass: 319 (M⁺)

PMR (Py-d₅): 3.46 (2H, t, J=8.1, H-6); 3.49 (3H, s, NCH₃); 4.65 (2H, t, J=8.1, H-5); 7.13 (1H, d, J=8.8, H-16); 7.21 (1H, dd, J=8.1, 6.3, H-18); 7.30 (2H, m, H-9, H-11); 7.38 (1H, d, J=2.2, H-2); 7.60 (2H, m, H-12, H-17); 8.33 (1H, m, H-10); 8.40 (1H, dd, J=8.1, 1.5, H-19); 11.85 (1H, d, J=2.2, H-1)

¹³C NMR (Py-d₅):

C-2	123.5 (d)	C-10	119.5 (d)	C-17	135.0 (d)
3	161.7 (s)	11	121.9 (d)	18	122.7 (d)
5	42.9 (t)	12	112.0 (d)	19	128.6 (d)
6	24.5 (t)	13	140.9 (s)	20	116.0 (s)
7	112.5 (s)	15	150.9 (s)	21	161.7 (s)
8	137.7 (s)	16	114.2 (d)	NCH ₃	30.5 (q)
9	119.3 (d)				

69. 1-HYDROXY-7,8-DEHYDRORUTECARPINE



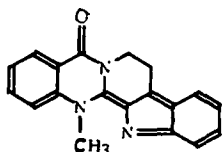
Vepris louisii [93]

C₁₈H₁₁N₃O₂: 301.3030

mp.: >340° (meth.)

UV: 230, 258, 294, 312, 363, 381, 402 (4.70, 4.69,
4.44, 4.30, 4.44, 4.64, 4.68)IR: 3360, 3270, 1690, 1670, 1640, 1610, 1580,
1550, 1490, 1380, 1330, 1245, 1145, 720Mass: 301 (M⁺, 100), 300 (4), 272 (4), 244 (10), 169 (7), 168 (46), 167 (7), 140 (13).PMR (DMSO-d₆): 7.20–7.80 (6H, m, H-Ar); 7.85 (1H, d, J=7.5, H-8); 8.18 (1H, dd, J=8, 2, H-4); 8.55 (1H, d, J=7.5, H-7); 9.36 (1H, br. s. OH); 12.36 (1H, br. s. NH)

70. DEHYDROEVODIAMINE



Evodia meliaefolia [95], E. rutaecarpa [109]

C₁₉H₁₅N₃O: 301.3466

mp.: 189–190°

{h-chl. 215–218°}

UV: 246, 313, 365 (4.51, 4.02, 4.71)

IR: 3440, 1700, 1610, 1550, 1430, 1380, 1280,
1260, 1235, 1210, 1105, 760, 750, 735

Mass: 301, 287, 286, 165, 156, 144.

PMR (DMSO-d₆): 3.33 (2H, t, J=6.7, H-6), 4.40 (3H, s, CH₃), 4.47 (2H, t, J=6.7, H-5); 7.27 (1H, t, J=8.5, H-10); 7.52 (1H, dt, J=1.2, 8.5, H-11); 7.72 (1H, d, J=8.5, H-12); 7.80 (1H, dt, J=1.2, 8.5, H-18); 7.88 (1H, d, J=8.5, H-8); 8.13 (1H, dt, J=1.2, 8.5, H-17); 8.19 (1H, d, J=8.5, H-16); 8.35 (1H, dd, J=1.2, 8.5, H-19); 12.70 (1H, s, H-1)

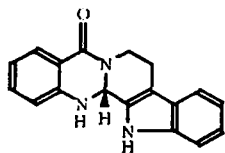
¹³C NMR (DMSO-d₆): [148]

C-21	158.2 (s)	C-18	128.6 (d)	C-20	118.7 (s)
3	149.9 (s)	19	127.7 (d)	16	118.4 (d)
13	141.4 (s)	8	123.3 (s)	12	113.5 (d)
15	139.6 (s)	10	121.6 (d)	5	42.0 (t)
17	136.7 (d)	9	121.5 (d)	22	40.8 (q)
7	130.4 (s)	2	120.0 (s)	6	18.5 (t)
11	128.8 (d)				

Pharm: hypotensive and negative chronotropic effects [203]

Uterine activity [204]. Vasodilator activity [223]

71. 13,14-DIHYDRORUTECARPINE

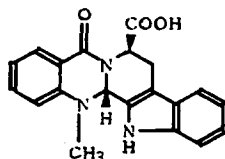


Evodia rutaecarpa [111]
 $C_{18}H_{15}N_3O$: 289.3356
 mp.: 214–216° (e-a) [111]; 226–228° [201]
 $[\alpha]_D^{25} -564^\circ$ (DMF)
 UV (meth.): 281, 290 (4.03, 3.91)

IR: 3330, 3280, 1633, 1618, 1609, 1425, 760, 750, 735
 Mass: 289 (M^+), 159, 143 (100), 130 [206]
 PMR (DMSO- d_6 , TMS): 2.65–3.40 (4H, m); 4.83 (1H, dt, $J=13.3$); 6.06 (1H, s); 6.70–7.60 (7H, m); 7.80 (1H, dd, $J=8.2$); 10.90 (1H, br. s)
 ^{13}C NMR [201]

C-21	163.5	C-8	126.5	C-16	115.0
15	147.1	18	125.7	12	111.5
13	136.0	11	121.7	7	109.1
17	133.2	9	119.8	3	63.7
2	130.6	10	118.9	5	40.5
19	127.9	20	115.8	6	19.9

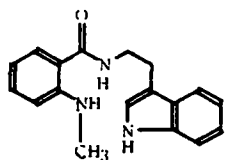
72. 7-CARBOXYEVODIAMINE



Evodia rutaecarpa [112]
 $C_{20}H_{17}N_3O_3$: 347.3714
 mp: amorph.
 $[\alpha]_D^{25} +441^\circ$ (chlf.)
 UV (acetonitrile): 268, 281, 289, 330
 IR (chlf.): 3475, 1745, 1665

Mass: 302, 300, 229, 201, 134
 PMR {COOCH $_3$ }: 2.46 (s, NCH $_3$); 3.22 ($J=16.6$, ABX); 3.55 ($J=16.1, 1.5$, ABX); 5.76 ($J=6.1, 1.5$, H-7, H-8); 6.22 (brs $W_{1/2}=3$, H-13b); 3.59 (s, COOCH $_3$); 7.10–7.70 (7H, H-Ar); 8.16 (1H, dd, $J=8.1, 1.5$, H-Ar); 8.41 (NH)
 Abs. conf.: 7S, 13b S

73. N-(2-METHYLAMINO BENZOYL)TRYPTAMINE

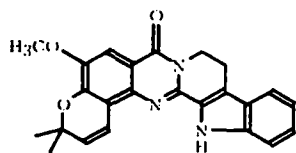


Evodia rutaecarpa [118]
 $C_{18}H_{19}N_3O$: 293.1528
 mp: 114–117° (bz.) [118]; 126–127° (toluene) [201]
 UV: 223, 259, 284, 292, 346 (4.40, 3.93, 3.47, 3.45, 3.55)

IR: 3450, 3272, 1633, 1583, 1519, 1280, 1220, 1170, 744, 733
 Mass: 293 (M^+)
 PMR (Py- d_5): 2.73 (3H, d, $J=4.9$, NHCH $_3$); 3.34 (2H, t, $J=7.3$, H-6); 4.00 (2H, q, $J=7.3$, H-5); 6.62 (1H, t, $J=7.3$, H-18); 6.71 (1H, d, $J=8.5$, H-16); 7.23 (1H, t, $J=7.3$, H-10); 7.29 (1H, dd, $J=8.6, 7.3$, H-11); 7.34 (1H, s, H-2); 7.36 (1H, dd, $J=8.5, 7.3$, H-17); 7.57 (1H, d, $J=8.6$, H-12); 7.90 (1H, d, $J=7.3$, H-19); 7.91 (1H, d, $J=7.3$, H-9); 8.32 (1H, bq, $J=4.9$, H-14); 9.01 (1H, br. t, $J=7.3$, H-4); 11.75 (1H, s, H-1) [118]
 ^{13}C NMR (Py- d_5): [118]

C-2	123.4	C-10	119.2	C-17	132.7
5	41.2	11	121.8	18	114.6
6	26.4	12	111.2	19	128.7
7	113.3	13	137.7	20	116.7
8	128.5	15	151.2	21	170.7
9	119.3	16	111.2	NHCH $_3$	29.6

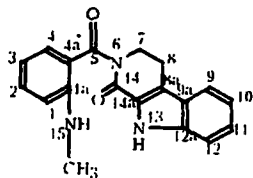
74. PARAENSINE



Euxylophora paraensis [119]
 $C_{24}H_{21}N_3O_3$: 399.4470
 mp.: 281–282° (bz.)
 UV (meth): 342, 358, 376 (4.32, 4.42, 4.29)
 IR (Nujol): 3310, 1650, 1640, 1600, 1550
 Mass: 399 (M^+), 384, 369, 358, 128, 115

PMR: 1.48 (6H, s, 2xCH $_3$); 3.42 (2H, t, $J=7$, CH $_2$ CH $_2$ N); 3.89 (3H, s, OCH $_3$); 4.51 (2H, t, $J=7$, CH $_2$ CH $_2$ N); 5.63 (d, $J=10$, H-olefin); 7.20–7.70 (H-Ar, H-olefin, NH); 7.53 (s, H-4)
 PMR (Py- d_5): 1.48, 3.10; 3.76; 4.62; 5.56; 7.05–7.85; 7.88

75. RHETSININE



Araliopsis tabouensis [114], *Zanthoxylum budrunga* [116], *Z. oxyphyllum* [117], *Z. rhetsa* [143]

$C_{19}H_{17}N_3O_2$: 319.3614

mp.: 185–187° (alc.-eth., dec.)

{h-chl. 258° (dec.)}

UV: 314

UV (alc. +NaBH₄): 283, 292 [114]

IR: 3400, 3250, 1690, 1675

Mass: 319 (M⁺) [116]; 186, 184, 157, 129, 105 [114]

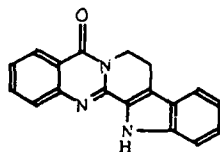
PMR: 3.32 (2H, t, J=6, NCH₂CH₂); 4.40 (3H, bs, NCH₃); 4.48 (2H, t, J=6, NCH₂CH₂); 7.28 (1H, t, J=10, H-3); 7.53 (1H, dt, J=10.2, H-2); 7.71 (1H, br.d, J=10, H-1); 7.81 (1H, dt, J=10.2, H-10); 7.89 (1H, bd, J=10, H-12); 8.14 (1H, dt, J=10.2, H-11); 8.20 (1H, bd, J=10, H-9); 8.36 (1H, dd, J=10.2, H-4); 12.60 (1H, s, NH)

¹³C NMR: [116]

C-1	127.6	C-7	42.0	C-11	121.5
1a	141.3	8	18.5	12	113.4
2	136.6	8a	118.7	12a	130.2
3	128.6	9	118.5	14	150.0
4	128.7	9a	123.2	14a	139.6
4a	120.0	10	121.5	NCH ₃	40.8
5	158.2				

HPLC: [202]

76. RUTECARPINE



Evodia meliaefolia [95], *E. rutaecarpa* [96], *Hortia arborea* [97], *H. badinii* [98], *H. regia* [99], *Zanthoxylum budrunga* [100], *Z. limonella* [101], *Z. pluviatile* [102]

$C_{18}H_{13}N_3O$: 287.1060

mp.: 259° (bz.-pet. eth.) [100]; 265–266° (chl. meth.) [102]

UV (meth) 276, 288, 330, 344, 361 [205]

IR: 3325, 1655

Mass: 287 (M⁺, 100), 286, 259, 258, 231, 229, 143.5, 143, 129 [206]

PMR: 3.23 (2H, t, J=7, ArCH₂); 4.60 (2H, t, J=7, CH₂N); 7.30–7.80 (8H, m, H-Ar); 8.30 (1H, s, NH) [207]

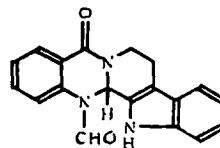
¹³C NMR [201]

C-21	159.9	C-19	125.9	C-9	119.3
15	146.8	18	125.9	10	119.3
3	143.6	16	125.3	7	117.3
13	138.1	8	124.1	12	112.1
17	133.7	11	124.1	5	40.5
2	126.4	20	120.2	6	18.8

HPLC: [202]

Pharm: vasodilator effect [223]

77. 14-FORMYLDIHYDRORUTECARPINE



Evodia rutaecarpa [111]

$C_{19}H_{15}N_3O_2$: 317.3456

mp.: 280–281° (meth.-DMF)

[α]_D²⁰: +260° (DMF)

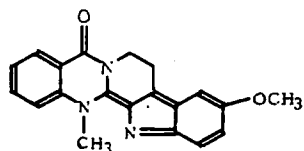
UV (meth.): 283, 291 (3.98, 3.91)

IR: 3342, 2918, 1690, 1608, 1493, 1472, 1413, 1378, 1327, 1318, 1302, 1282, 1240, 868, 771, 761, 747, 702

Mass: 318 (4), 317 (M⁺, 16), 289 (11), 288 (52), 287 (100), 286 (53), 170 (22), 169 (62), 148 (11), 147 (9), 146 (18), 143 (13)

PMR (DMSO-d₆, TMS): 2.60–3.70 (3H, m); 4.72 (1H, dd, J=12.5, 5); 6.80–7.70 (8H, m); 7.92 (1H, d, J=8); 9.10 (1H, s); 11.05 (1H, br.s)

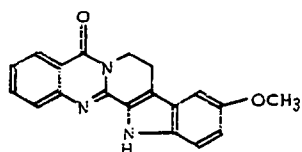
¹³ C NMR [201]					
C-15	163.2	C-18	126.0	C-10	117.8
21	163.1	8	125.8	12	111.4
NCHO	162.0	20	119.8	7	110.8
13	136.8	11	119.5	3	61.8
17	133.0	16	119.2	5	43.6
2	131.2	9	118.0	6	19.0
19	126.8				



Hortia arborea [103], H. braziliana [110]
 $C_{20}H_{17}N_3O_2$: 331.1744
 mp.: 208° (bz.)
 {h-chl. 243° (alc. water, dec.) m-i. 209° (alc. water, dec.)}

UV (acetonitrile): 411 (47300) [97]
 Pharm: hypotensive action [110]

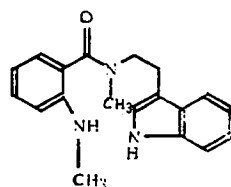
78. HORTIAMINE



Hortia arborea [103], H. badini [98], H. longitolia [104]
 $C_{19}H_{15}N_3O_2$: 317.3456
 mp.: 252° (e-a)
 IR: 3338, 1648, 1611, 1491, 1461, 1394, 1213, 1162, 1098, 1026, 798, 761 [201]

Mass: 318 (19), 317 (M⁺, 100), 302 (23), 284 (22) [201]

79. HORTIACINE



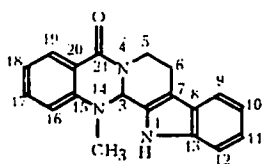
Evodia rutaecarpa [115]
 $C_{19}H_{21}N_3O$: 307.3940
 mp.: 208–209° (bz.)
 UV: 221, 250, 284, 291, 311
 IR: 3350, 3200, 1620, 1600, 1580, 1518, 1285, 1235, 1120, 750
 Mass: 307 (M⁺)

PMR: 2.66 (3H, d, J=4.4, NHCH₃); 3.00 (3H, br.s, NCH₃); 3.21 (2H, m, H-6); 3.86 (2H, m, H-5); 5.71 (1H, m, 14-NH); 6.71 (1H, d, J=8.8, H-9); 6.75 (1H, d, J=7.3, H-18); 7.24–7.36 (6H, m, H-2, H-10, H-11, H-12, H-17, H-19); 7.58 (1H, d, J=7.3, H-9); 11.86 (1H, s, 1-NH)

¹³ C NMR					
C-2	123.4	C-11	121.8	C-17	130.6
6	24.1	12	112.0	18	115.8
8	128.3	13	137.6	19	128.0
9	119.1	15	147.6	21	171.0
10	119.2	16	110.9	NHCH ₃	30.0

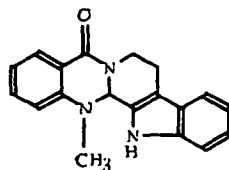
80. EVODIAMIDE

81. (±)-EVODIAMINE



Zanthoxylum rhetsa [113]
 $C_{19}H_{17}N_3O$: 303.3624
 mp.: 277°
 HPLC: [223]

82. (+)-EVODIAMINE



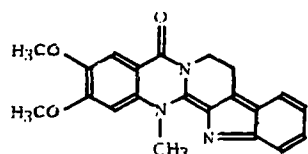
Araliopsis tabouensis [114], Evodia meliaefolia [95],
E. rutaecarpa [96], Zanthoxylum rhetsu [113]
C₁₉H₁₇N₃O: 303.3624
mp.: 278°
[α]_D²⁵: +352°
UV: 314;
UV (alc. -NaBH₄): 283, 292

IR: 3400, 3250, 1690, 1675
Mass: 303 (M⁺, 100), 302, 274, 170, 169, 151.5, 143, 134 [206]
PMR: 2.53 (3H, s, NCH₃); 5.90 (1H, s, H-3); 7.30-7.80 (8H, m, H-Ar); 8.20 (1H, s, NH) [207]
¹³C NMR [201]

C-21	164.1	C-8	125.8	C-12	111.5
15	148.7	11	121.8	7	111.4
13	136.3	9	120.2	3	69.7
17	133.3	20	119.2	5	39.5
2	130.3	10	118.8	NCH ₃	36.4
18	127.4	16	118.1	6	19.4
19	127.3				

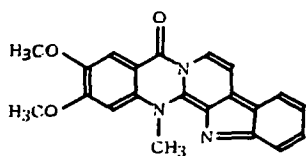
HPLC: [202, 203]
Pharm: vasodilator effect [223], Antianoxic effect [224]

83. EUXYLOPHORINE A



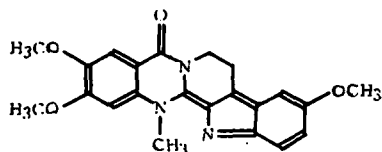
Euxylophora paraensis [105]
C₂₁H₁₉N₃O₃: 361.3982
mp.: 227-230° (bz.)
UV (acetonitrile): 253, 402 (4.44, 4.60)
IR (Nujol) 1670, 1656, 1618, 1603, 1545
PMR (Py-d₅): 3.30 (2H, t); 3.90 (s, OCH₃); 3.96 (s, OCH₃); 4.75 (2H, t); 5.23 (s, NCH₃);
7.20-8.20 (6H, m, H-Ar)

84. EUXYLOPHORINE B



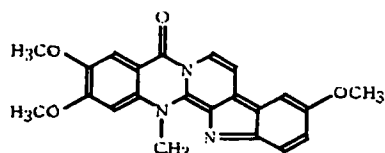
Euxylophora paraensis [106]
C₂₁H₁₇N₃O₃: 359.3824
mp.: 268-271° (chlf., dec.)
{h-chl. 270-280° (meth., dec.)
UV (acetonitrile) 278, 352 (4.47, 4.70)
UV (h-chl., acetonitrile): 297, 335, 346, 405 (4.28, 4.18, 5.43, 3.85)
IR (Nujol) 1690, 1618, 1605, 1555, 1510
IR {h-chl., Nujol}: 3320, 1710, 1615, 1580
Mass: 359 (M⁺)
PMR (CD₃COOD+ chlf.): 4.22 (3H, s, OCH₃); 4.30 (3H, s, OCH₃); 4.81 (3H, br.s, NCH₃); 7.37 (1H, s, H-1); 8.05 (1H, s, H-4); 7.40-8.30 (4H, m, H-Ar); 8.32 (1H, d, J=9, H-8); 9.34 (1H, d, J=9, H-7); 11.70 (1H, br.s., NH).

85. EUXYLOPHORINE C



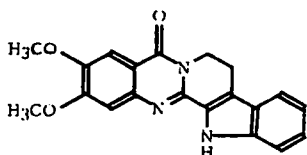
Euxylophora paraensis [107]
C₂₂H₂₁N₃O₄: 391.4240
mp.: 207-209° (bz., dec.)
UV (acetonitrile) 303, 408 (4.14, 4.40)
IR: 1665, 1620, 1555
Mass 391 (M⁺)
PMR: 3.40 (2H, t, J=7, H-8); 4.02 (3H, s, OCH₃); 4.12 (3H, s, OCH₃); 4.15 (3H, s, OCH₃); 4.50 (3H, br.s, W_{1/2}=3, NCH₃); 4.75 (2H, t, J=7, H-7); 7.20-7.60 (4H, m, H-Ar); 7.85 (1H, s, H-4); 10.10 (1H, br.s., NH)

86. EUXYLOPHORINE D



Euxylophora paraensis [107]
 $C_{22}H_{19}N_3O_4$: 389.4082
 mp.: 256–260° (bz., dec.)
 UV (acetonitrile) 277, 360 (4.33, 4.65)
 Mass: 389 (M^+)

87. EUXYLOPHORICINE A



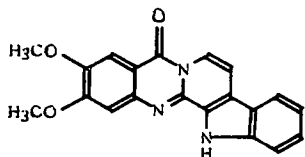
Euxylophora paraensis [105]
 $C_{20}H_{17}N_3O_3$: 347.3714
 mp.: 295–298° (chlf.-meth) [105]; 294–295° [201]
 UV (meth.) 255, 337, 353, 360 (4.50, 4.49, 4.54, 4.43)
 IR (Nujol): 3300–3200, 1650, 1615, 1590

PMR: 3.10 (t): 3.80 (s, OCH_3): 3.85 (s, OCH_3): 4.60 (t): 7.20–7.90 (m, NH, H-Ar)

^{13}C NMR [201]

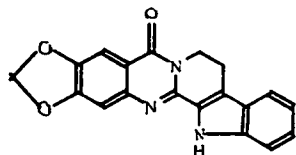
159.7 (s)	127.3 (s)	112.4 (d)
154.5 (s)	125.0 (s)	107.1 (d)
148.2 (s)	124.3 (d)	106.0 (d)
144.0 (s)	119.7 (2d)	55.8 (2q)
143.2 (s)	116.8 (s)	37.6 (t)
138.5 (s)	113.7 (s)	18.9 (t)

88. EUXYLOPHORICINE B



Euxylophora paraensis [105]
 $C_{20}H_{15}N_3O_3$: 345.3556
 mp.: 310–312° (chlf.-meth.)
 UV: 256, 294, 304, 330, 353, 372, 392 (4.54, 4.40, 4.53, 4.37, 4.27, 4.45, 4.51)
 IR (Nujol) 3350, 1656, 1634, 1600, 1575
 PMR (CD_3COOD): 8.20, 9.13 (J=7, H-7, H-8)

89. EUXYLOPHORICINE C



Euxylophora paraensis [106]
 $C_{19}H_{13}N_3O_3$: 331.3288
 mp.: 310–312° (sp.)
 UV (acetonitrile): 252, 337, 350, 368 (4.55, 4.50, 4.53, 4.36)

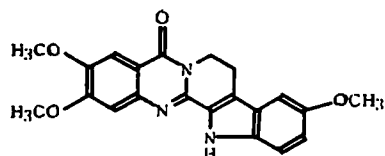
IR (Nujol): 3350, 1655, 1630, 1600, 1550, 940 [106]

IR: 3345, 1653, 1631, 1598, 1471, 1331, 1228, 1131, 729 [201]

Mass 331 (M^+)

PMR (CD_3COOD +chlf.): 3.58 (2H, t, J=7, H-8); 4.82 (2H, t, J=7, H-7); 6.28 (2H, s, OCH_2O): 7.28 (s, H-1); 7.72 (1H, s, H-4); 7.20–7.80 (4H, m, H-Ar); 10.60 (1H, br.s, NH)

90. EUXYLOPHORICINE D



Euxylophora paraensis [107]
 $C_{21}H_{19}N_3O_4$: 377.3972
 mp.: 293–295° (chlf.) [107]; 291–292° [201]
 UV (acetonitrile) 252, 341, 356, 374 (4.44, 4.45, 4.48, 4.32)

IR: 3450, 1670, 1620, 1600 [107]

IR: 3365, 1652, 1612, 1492, 1460, 1292, 1230, 1097, 818

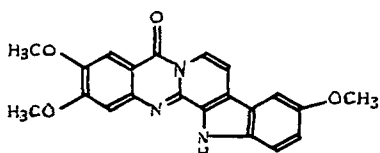
Mass 377 (M^+)

PMR : 3.43 (2H, t, J=7, H-8); 4.03 (3H, s, OCH₃); 4.10 (6H, s, 2xOCH₃); 4.78 (2H, t, J=7, H-7); 7.20-7.60 (4H, m, H-Ar); 7.78 (1H, s, H-4); 7.95 (1H, br.s, NH)

¹³C NMR: [201]

159.7 (s)	133.6 (s)	113.2 (d)
154.4 (s)	127.5 (s)	105.8 (d)
153.6 (s)	125.0 (s)	100.3 (d)
148.0 (s)	116.3 (s)	55.6 (2q)
144.0 (s)	115.4 (d)	38.0 (d)
143.2 (s)	113.5 (s)	18.9 (t)

91. EUXYLOPHORICINE E



Euxylophora paraensis [107]

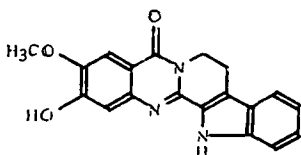
C₂₁H₁₇N₃O₄: 375.3814

mp.: 290° (chlf.)

UV(chlf.): 255, 289, 292, 307, 355, 376, 398
(4.65, 4.55, 4.48, 4.54, 4.57, 4.62, 4.64)

Mass 375 (M⁺)

92. EUXYLOPHORICINE F



Euxylophora paraensis [108]

C₁₉H₁₅N₃O₃: 333.3446

mp.: 226° (bz.-pet.eth.)

UV (meth.) 247, 337, 347, 364 (4.50, 4.48, 4.51, 4.43)

UV (meth. +NaOH): 304

UV (meth. +HCl): 372

IR: 3300

Mass: 333 (M⁺, 100), 332 (22), 318 (15), 303.6, 264.4, 166.5 (13)

PMR (CD₃COOD-chlf.): 3.53 (t, CH₂CH₂N); 4.90 (t, J=7.0, CH₂CH₂N); 4.20 (s, OCH₃); 7.20-7.80 (5H, m, H-Ar); 7.92 (1H, s, H-4)

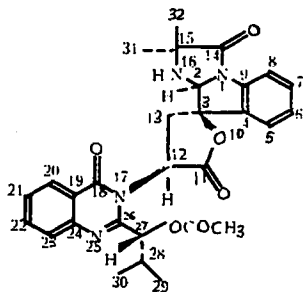
IV. Tryptoquivalines

Tryptoquivaline and its analogs are products of the vital activity of three species of ascomycetous fungi (Ascomycetes): *Aspergillus clavatus*, *Aspergillus fumigatus*, and *Corynascus setosus*. Up to the present time, two groups of workers, from the USA and Japan, have isolated 19 substances of this nature.

They are all fairly complex heterocyclic systems consisting of six rings, one of which is a spiro- γ -lactone. The latter is attached to the indole part of the molecule in position 3. The indole component, in its turn, is linked with a saturated imidazole ring by an N₁-C₂ bond. The quinazoline unit is bound to the spiro- γ -lactone ring by an N₁₇-C₁₂ bond. In all these structures, the presence of three carbonyl functions at carbon atoms 11, 14, and 18 is obligatory. The N₁₆ nitrogen may have the following substituents: hydrogen, hydroxyl, methoxycarbonyl. The quinazoline part of the molecule frequently contains a grouping of one of three types: CH(OH)-CH(CH₃)₂, CH(OAc)-CH(CH₃)₂, CO-CH(CH₃)₂. Finally, in position 15 there are one or two methyl groups. In addition to the complex structure, which was revealed mainly by the XSA method, the tryptoquivalines possess four or five asymmetric centers. These are the C₂, C₃, C₁₂, C₁₅, and C₂₇ carbon atoms. The absolute configurations of these centers were also determined for nortryptoquivaline by the XSA method [124].

It is interesting to note that in the microorganism *Aspergillus fumigatus* tryptoquivalines are produced together with other quinazoline derivatives — fumiquinazolines A, B, and C, which are discussed below.

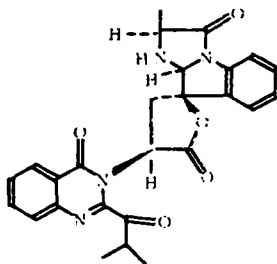
93. DEOXYNORTRYPTOQUIVALINE



Aspergillus clavatus [120]
 $C_{28}H_{28}N_4O_6$; 516.2009
 mp.: 158–160° (eth.)
 $[\alpha]_D^{20} -70^\circ$ (chlif.)
 UV 228, 233, 254, 268, 278, 305, 317 (43900,
 40100, 15600, 11700, 10500, 4100, 3300)
 IR (chlif.): 3360, 2975, 2935, 2880, 1790, 1724,
 1676, 1607, 1483
 Mass 516 (M^+ , 100)

PMR 1.00 (3H, d, J=7); 1.15 (3H, d, J=7); 1.55 (3H, d, J=7); 2.16 (3H, s); 2.58 (1H, m); 2.87
 (1H, dd, J=10, 13); 3.07 (1H, dd, J=10, 13); 4.12 (1H, q, J=7); 5.22 (1H, s); 5.55 (1H, d,
 J=9); 5.65 (1H, t, J=10); 7.02–7.74 (7H, m); 8.14 (1H, m)

94. DEOXYNORTRYPTQUIVALONE

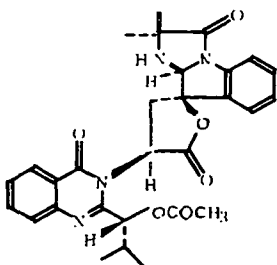


Aspergillus clavatus [120]
 $C_{26}H_{24}N_4O_5$; 472.1747
 mp.: 192–193° (eth.)
 $[\alpha]_D^{20} +171^\circ$ (chlif.)
 UV 232, 288, 326 (32400, 9250, 6250)
 IR (chlif.): 3360, 2980, 2935, 2880, 1790, 1705,
 1686, 1610, 1588, 1484, 1469
 Mass 472 (M^+)

PMR: 1.26 (3H, d, J=7); 1.30 (3H, d, J=7); 1.56 (3H, d, J=7); 3.02 (1H, dd, J=10, 13); 3.32
 (1H, dd, J=10, 13); 4.08 (1H, m, J=7); 4.12 (3H, q, J=7); 5.36 (1H, s); 5.48 (1H, t, J=10);
 7.04–7.84 (7H, m); 8.24 (1H, m)

HPLC

95. DEOXYTRYPTOQUIVALINE

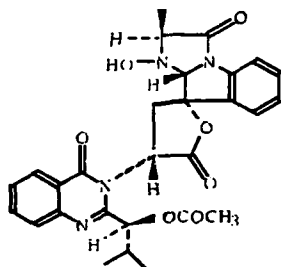


Aspergillus clavatus [120]
 $C_{29}H_{30}N_4O_6$; 530.5768
 mp.: 150–152° (meth. chl.-hx.)
 $[\alpha]_D^{20} -57^\circ$ (chlif.)
 UV 227, 232, 252, 267, 278, 304, 318 (44500, 41900,
 18500, 12000, 10300, 3300, 2700)
 IR (chlif.): 3360, 3310, 2980, 2935, 2875, 1790, 1720,
 1676, 1604, 1483, 1469
 Mass 530 (M^+ , 100)

PMR: 1.04 (3H, d, J=7); 1.20 (3H, d, J=7); 1.53 (6H, s); 2.16 (3H, s); 2.54 (1H, m); 3.06 (2H,
 d, J=10); 5.24 (1H, s); 5.52 (1H, d, J=9); 5.65 (1H, t, J=10); 7.11–7.74 (7H, m); 8.20 (1H,
 m)

HPLC

96. NORTRYPTOQUIVALINE



Aspergillus clavatus [120], *A. fumigatus* [124]
 $C_{28}H_{28}N_4O_7$; 532.1956
 mp.: 256–258° (meth. chl.-hx.)
 $[\alpha]_D^{20} -170^\circ$ (chlif.)
 UV 228, 233, 254, 267, 279, 306, 319 (43600,
 42000, 18700, 11900, 10200, 4500, 3500)
 IR (chlif.): 3490, 2980, 2940, 2880, 1790, 1728,
 1670, 1610, 1485, 1471, 1410
 Mass 532 (M^+)

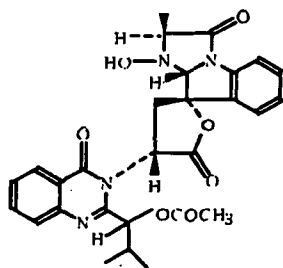
PMR 1.02 (3H, d, J=7); 1.16 (3H, d, J=7); 1.58 (3H, d, J=7); 2.16 (3H, s); 2.57 (1H, m); 2.94
 (1H, dd, J=10, 13); 3.18 (1H, dd, J=10, 13); 4.28 (1H, q, J=7); 5.10 (1H, s); 5.54 (1H, d,
 J=9); 5.65 (1H, t, J=10); 7.01–7.79 (7H, m); 8.12 (1H, m)

XSA: [124]

Abs. conf. 2S, 3S, 12R, 15S, 27S [124]

HPLC

Pharm: antibacterial activator



Corynascus setosus [208]

C₂₈H₂₉N₄O₇; 533.2023

mp.: 236–238° (meth.)

[α]_D²⁰ +196° (chlf.)

UV (meth.) 209, 290, 307, 318 (4.64, 3.80, 3.53, 3.43)

IR (chlf.): 3500, 3000, 1780, 1720, 1660, 1600

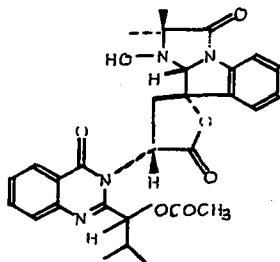
Mass 533.2023 (M⁺)

PMR 5.23 (s, H-2); 7.41 (bd, J=7.6, H-5); 7.28 (td, J=7.6, 1.1, H-6); 7.49 (td, J=7.6, 1.1, H-7); 7.63 (br.d, J=7.6, H-8); 5.87 (t, J=10.1, H-12); 3.09 (dd, J=13.3, 9.5, H-13); 3.15 (dd, J=13.3, 10.7, H-13); 4.35 (q, J=7.2, H-15); 7.16 (s, 16-OH); 8.22 (dd, J=8.4, 1.5, H-20); 7.55 (td, J=8.4, 1.3, H-21); 7.84 (td, J=8.4, 1.5, H-22); 7.79 (br.d, J=8.4, H-23); 5.66 (d, J=9.8, H-27); 2.68 (m, H-28); 0.98 (3H, d, J=6.6, H-29); 1.16 (3H, d, J=6.6, H-30); 2.21 (3H, s, H-33); 1.59 (3H, d, J=7.2, H-34)

¹³C NMR

C-2	89.9 (d)	C-13	34.1 (t)	C-24	146.5 (s)
3	83.9 (s)	14	169.2 (s)	26	151.5 (s)
4	133.7 (s)	15	67.9 (d)	27	79.0 (d)
5	123.7 (d)	18	161.8 (s)	28	31.8 (d)
6	125.7 (d)	19	120.3 (s)	29	19.0 (q)
7	131.9 (d)	20	126.7 (d)	30	18.9 (q)
8	116.1 (d)	21	128.3 (d)	32	170.5 (s)
9	137.8 (s)	22	135.5 (d)	33	20.8 (q)
11	169.3 (s)	23	128.0 (d)	34	11.0 (q)
12	54.6 (d)				

Abs. conf. 27R



Corynascus setosus [208]

C₂₉H₃₁N₄O₇; 547.2183

mp.: 225–227° (meth.)

[α]_D²⁰ -138° (chlf.)

UV (meth.) 209, 289, 306, 318 (4.67, 3.83, 3.54, 3.44)

IR (chlf.): 3500, 3000, 1790, 1735, 1675, 1610

Mass 547.2183 (M⁺)

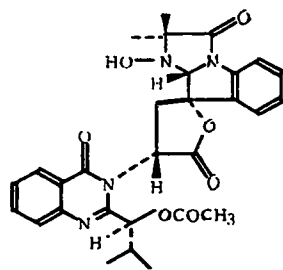
PMR 5.05 (s, H-2); 7.41 (br.d, J=7.6, H-5); 7.28 (td, J=7.6, 1.1, H-6); 7.48 (td, J=7.6, 1.1, H-7); 7.64 (br.d, J=7.6, H-8); 5.88 (t, J=10.0, H-12); 3.12 (dd, J=13.5, 9.7, H-13); 3.14 (dd, J=13.5, 10.1, H-13); 6.99 (s, 16-OH); 8.25 (dd, J=7.2, 1.3, H-20); 7.55 (td, J=7.2, 1.2, H-21); 7.84 (td, J=7.2, 1.3, H-22); 7.79 (br.d, J=7.2, H-23); 5.65 (d, J=10.1, H-27); 2.69 (m, H-28); 1.06 (3H, d, J=6.8, H-29); 1.17 (3H, d, J=6.8, H-30); 2.22 (3H, s, H-33); 1.49 (3H, s, H-34); 1.50 (3H, s, H-35)

¹³C NMR

C-2	87.6 (d)	C-13	34.6 (t)	C-26	151.5 (s)
3	83.9 (s)	14	169.4 (s)	27	78.9 (d)
4	133.7 (s)	15	71.4 (s)	28	31.8 (d)
5	123.7 (d)	18	161.7 (s)	29	18.9 (q)
6	125.6 (d)	19	120.3 (s)	30	18.9 (q)
7	131.9 (d)	20	126.8 (d)	32	170.6 (s)
8	116.1 (d)	21	128.2 (d)	33	20.8 (q)
9	138.1 (s)	22	135.5 (d)	34	16.9 (q)
11	169.4 (s)	23	128.0 (d)	35	22.8 (q)
12	54.7 (d)	24	146.5 (s)		

Abs. conf. 27R

99. TRYPITOQUIVALINE



Aspergillus clavatus [125], *A. fumigatus* [121]
 $C_{29}H_{30}N_4O_7$: 546.2114
 mp : 153–155°
 $[\alpha]_D^{25}$ -142° (chlf.)
 UV 228, 275, 305, 317 (37000, 8550, 3800, 3040)
 IR (chlf.), 3520, 1790, 1735, 1680, 1615
 Mass 546 (M^+)

PMR 1.03 (3H, d, J=7); 1.17 (3H, d, J=7); 1.50 (3H, s); 1.52 (3H, s); 2.19 (3H, s); 2.63 (1H, m); 3.10 (1H, d, J=10); 3.15 (1H, d, J=10); 3.63 (1H); 4.04 (1H); 5.00 (1H, c); 5.61 (1H, d, J=9); 5.70 (1H, t, J=10); 7.12–7.90 (7H, m); 8.22 (1H, d, J=8) [125]

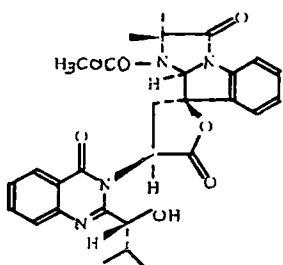
XSA {p-bromophenylurethane deriv.} [125]

Abs. conf: 27S [208]

HPLC [120]

Pharm: anticonvulsive activity

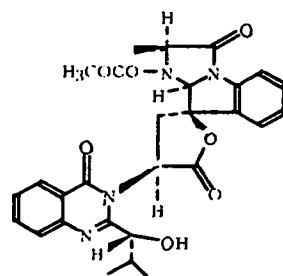
100. TRYPITOQUIVALINE C



Aspergillus fumigatus [121]
 $C_{29}H_{30}N_4O_7$: 546.5758
 mp : 215–217° (meth., dec.)
 $[\alpha]_D^{25}$ -168° (chlf.)
 UV (meth.) 228, 233, 255, 280, 307, 320 (39800, 38000, 21100, 14200, 4200, 2700)
 IR: 3490, 1760, 1745, 1665, 1600
 Mass 546 (M^+)

PMR 1.04 (3H, d, J=6); 1.14 (3H, d, J=6); 1.48 (3H, s); 1.50 (3H, s); 2.17 (3H, s); 3.03 (1H, dd, J=14, 10); 3.20 (1H, dd, J=14, 10); 2.67 (1H, m); 4.99 (1H, s); 5.59 (1H, d, J=9); 5.67 (1H, t, J=10); 7.08 (1H, s); 7.33–7.93 (7H, m); 8.22 (1H, d, J=8)

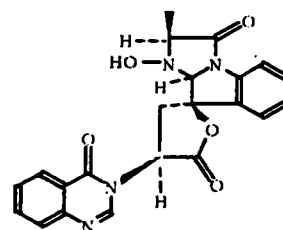
101. TRYPITOQUIVALINE D



Aspergillus fumigatus [121]
 $C_{28}H_{28}N_4O_7$: 532.5490
 mp.: 224–225° (meth., dec.)
 $[\alpha]_D^{25}$ -115° (chlf.)
 UV (meth.) 227, 233, 257, 277, 305, 319 (48100, 45200, 21500, 14500, 4800, 3300)
 IR 3430, 1778, 1732, 1660, 1610
 Mass 532 (M^+)

PMR: 0.97 (3H, d, J=7); 1.09 (3H, d, J=7); 1.56 (3H, d, J=7); 2.14 (3H, s); 2.57 (1H, m); 2.93 (1H, dd, J=14.9); 3.19 (1H, dd, J=14.9); 4.32 (1H, q, J=7); 5.10 (1H, s); 5.61 (1H, d, J=8); 5.74 (1H, t, J=9); 7.26 (1H, s); 7.11–7.99 (7H, m); 8.24 (1H, d, J=8)

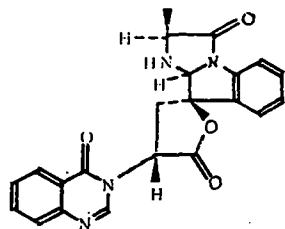
102. TRYPITOQUIVALINE E



Aspergillus fumigatus [122]
 $C_{22}H_{18}N_4O_5$: 418.1277
 mp : 257° (ac., dec.)
 $[\alpha]_D^{25}$ -257° (chlf.)
 UV (meth.) 225.5, 232, 254, 265.5, 275.5, 291, 303, 315 (32300, 29800, 16000, 11700, 8400, 3500, 3000, 2600)
 IR 3430, 1780, 1740, 1732, 1677, 1658, 1614
 Mass 418 (M^+ , 100), 402 (94), 356 (22), 254 (37), 215 (86)

PMR (Py- d_5): 1.63 (d, J=7, CH_3); 3.38, 3.56 (dd, J=13, 10, CH_2); 4.26 (q, J=7, CH); 5.42 (s, CH); 6.48 (t, J=10, CH); 6.83–7.80 (7H, m, H-Ar); 8.17 (1H, d, J=8, =CH-); 8.59, 8.66 (1H, s, =CH-); 10.42 (br. s, NOH)

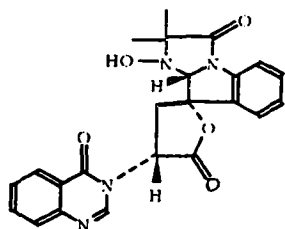
103. TRYPTOQUIVALINE F



Aspergillus fumigatus [122]
 $C_{22}H_{18}N_4O_4$: 402.1328
 mp.: 277° (meth., dec.)
 $[\alpha]_D^{20} -109^\circ$ (chlf.)
 UV (meth.) 226, 232, 255, 265, 276, 290, 303, 315
 (33400, 31200, 12400, 11100, 8100, 3100,
 2800, 2200)
 IR: 3365, 1775, 1725, 1664, 1606
 Mass 402 (M^+ , 50), 374 (22), 215 (47), 146 (100)

PMR (Ac.): 1.68 (d, J=7, 31-CH₃, 32-CH₃); 2.15 (s, Ac); 3.19, 3.33 (dd, J=13.9, 13-CH₂); 4.44 (q, J=7, 15-CH); 5.40 (t, J=9, 12-CH); 5.65 (s, 2-CH); 7.12-8.07 (7H, m, H-Ar); 8.04 (1H, s, H-26); 8.21 (1H, d, J=7, H-20)

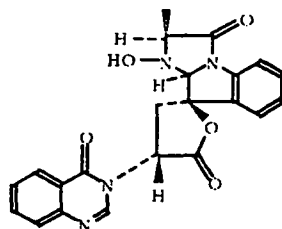
104. TRYPTOQUIVALINE G



Aspergillus fumigatus [122]
 $C_{23}H_{20}N_4O_5$: 432.1433
 mp: 240-241.5° (ac., dec.)
 $[\alpha]_D^{20} -215^\circ$ (ac.)
 UV (meth.) 226, 232, 253, 265, 275, 291, 302, 315
 (34300, 31700, 17400, 11800, 8200, 3700,
 3000, 2500)
 IR: 3470, 1778, 1738, 1662, 1610.
 Mass 432 (M^+ , 53); 414 (12), 386 (16), 242 (68),
 229 (100), 228 (65)

PMR (Py-d₅): 1.50 (s, 31-CH₃); 1.62 (s, 32-CH₃); 3.43, 3.64 (dd, J=14, 10, 13-CH₂); 5.30 (s, 2-CH); 6.56 (t, J=10, 12-CH); 6.96-7.96 (7H, m, H-Ar); 8.23 (1H, d, J=8, H-20); 8.66 (1H, s, H-26); 10.64 (br. s, NOH)

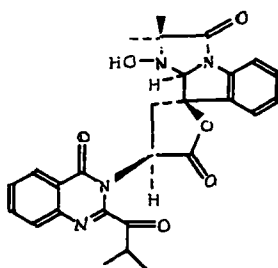
105. TRYPTOQUIVALINE H



Aspergillus fumigatus [122]
 $C_{22}H_{18}N_4O_5$: 418.1277
 mp 274° (meth., decomp.)
 $[\alpha]_D^{20} -155^\circ$ (ac.)
 UV (meth.) 226, 232, 255, 266, 276, 291, 303, 315
 (33100, 30900, 16600, 11300, 8500, 3600,
 3100, 2500)
 IR 3430, 1780, 1742, 1734, 1667, 1609
 Mass 418 (M^+ , 20), 400 (7), 215 (47), 146 (100)

PMR (Py-d₅): 1.45 (d, J=8, 31-CH₃, 32-CH₃); 2.96, 3.60 (dd, J=13, 10, 13-CH₂); 3.98 (q, J=8, 15-CH); 5.25 (s, 2-CH); 5.76 (t, J=10, 12-CH); 6.68-7.88 (7H, m, H-Ar); 8.11 (1H, d, J=8, H-20); 8.38, 8.41 (1H, s, H-26); 10.60 (br. s, NOH)

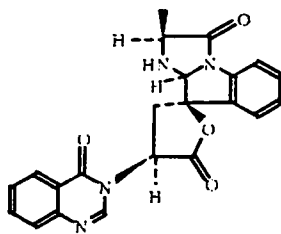
106. TRYPTOQUIVALINE I



Aspergillus fumigatus [122]
 $C_{27}H_{26}N_4O_6$: 502.1852
 mp: 232-235.5° (meth. chl.-meth, dec.)
 $[\alpha]_D^{20} +239^\circ$ (chlf.)
 UV (meth.) 235, 250, 292, 321 (31700, 21400, 9600,
 6100)
 IR: 3480, 1780, 1732, 1710, 1675, 1609.
 Mass 502 (M^+ , 28), 486 (19), 252 (20), 242 (32),
 229 (100)

PMR 1.22 (d, J=7, 29-CH₃); 1.28 (d, J=7, 30-CH₃); 1.49 (6H, s, 31-CH₃, 32-CH₃); 3.06, 3.39 (dd, J=14, 10, 13-CH₂); 4.07 (q, J=7, 28-CH); 4.99 (s, 2-CH); 5.47 (t, J=10, 12-CH); 7.00-7.94 (7H, m, H-Ar); 8.24 (1H, d, J=7, H-20); 7.01 (s, NOH)

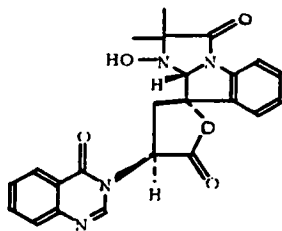
107. TRYPTOQUIVALINE J



Aspergillus fumigatus [122]
 $C_{22}H_{18}N_4O_4$: 402.1328
 mp.: 254–258° (ac.-meth., water)
 $[\alpha]_D^{25} +135^\circ$ (chl.f.)
 UV (meth.): 225.5, 231, 253, 264, 275, 290, 302, 310
 (41100, 38000, 16200, 12800, 9700, 4200,
 3900, 3100)
 IR: 3375, 1780, 1713, 1670, 1610
 Mass: 402 (M^+ , 59), 215 (24), 202 (23), 174 (71),
 44 (100)

PMR(DMSO- d_6): 1.41 (d, $J=7$, 31- CH_3 , 32- CH_3); 3.08, 3.10 (d, $J=10$, 13- CH_2); 3.87 (q, $J=7$, 15- CH); 5.41 (bd, $J=6$, 2- CH); 6.01 (t, $J=10$, 12- CH); 7.20–8.02 (7H, m, H-Ar); 8.17 (1H, d, $J=8$, H-20); 8.49 (1H, s, H-26); 3.76 (NH)

108. TRYPTOQUIVALINE L

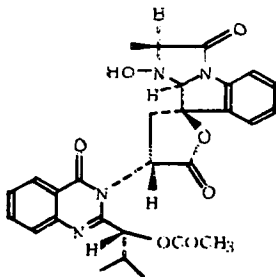


Aspergillus fumigatus [123]
 $C_{23}H_{20}N_4O_5$: 432.1457
 mp.: 265–268° (ac.)
 $[\alpha]_D^{25} -229^\circ$ (DMSO)
 UV (meth.): 216, 226, 231, 252, 264, 274, 290,
 302, 315 (33900, 33000, 30800, 17400,
 12300, 8400, 3700, 3000, 2400)
 IR: 3230, 1784, 1749, 1670, 1616, 1485, 1260,
 1200

Mass: 432 (M^+ , 76), 286 (13), 269 (15), 256 (16), 242 (28), 229 (100); 199 (16), 188 (27), 173
 (13), 147 (39), 130 (26)

PMR(DMSO- d_6): 1.26 (3H, s); 1.36 (3H, s); 3.03 (1H, dd, $J=13, 10$); 3.43 (1H, dd, $J=13, 10$);
 5.21 (1H, s); 5.57 (1H, t, $J=10$); 7.24–8.00 (7H, m); 8.23 (H, d, $J=8$); 8.53 (1H, s); 8.74
 (1H, s)

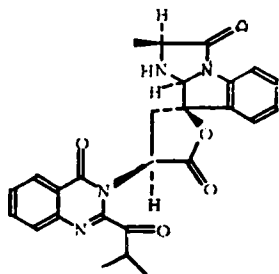
109. TRYPTOQUIVALINE M



Aspergillus fumigatus [123]
 $C_{26}H_{28}N_4O_7$: 532.1932
 mp.: 157–164° (meth.-water)
 $[\alpha]_D^{25} -154^\circ$ (chl.f.)
 UV (meth.): 228, 232, 255, 278, 305, 317 (32700,
 30700, 15400, 9300, 2900, 2700)
 IR: 3400, 1788, 1725, 1678, 1600, 1480, 1464, 1210
 Mass: 532 (M^+ , 100), 261 (33), 255 (23), 215 (50),
 214 (22), 201 (56), 130 (19)

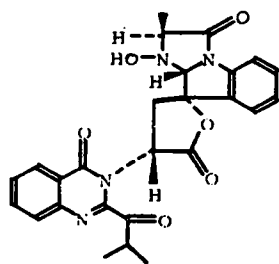
PMR: 0.92 (3H, d, $J=6$); 1.04 (3H, d, $J=6$); 1.55 (3H, d, $J=7$); 2.16 (3H, s); 2.68 (1H, dd, $J=11$,
 10); 2.74 (1H, m); 3.65 (1H, dd, $J=11, 10$); 4.15 (1H, q, $J=7$); 5.22 (1H, s); 5.54 (1H, d,
 $J=10$); 5.87 (1H, t, $J=10$); 6.86 (1H, br.s); 7.12–7.88 (7H, m); 8.24 (1H, d, $J=8$)

110. TRYPTOQUIVALINE N



Aspergillus fumigatus [123]
 $C_{26}H_{24}N_4O_5$: 472.1770
 mp.: 193–197° (meth.)
 $[\alpha]_D^{25} +127^\circ$ (DMSO)
 UV (meth.): 232, 251, 291, 320 (32300, 17900, 9000,
 6100)
 IR: 3360, 1780, 1722, 1705, 1680, 1607, 1481, 1250
 Mass: 472 (M^+ , 100), 444 (22), 256 (40), 243 (17),
 228(48), 217 (44), 199 (17), 186 (20), 174
 (44), 158 (24), 146 (38), 130 (35)

PMR: 1.26 (3H, d, $J=7$); 1.30 (3H, d, $J=7$); 1.55 (3H, d, $J=7$); 3.03 (1H, dd, $J=13.10$); 3.05
 (1H); 3.31 (1H, dd, $J=13.10$); 4.08 (1H, m); 4.12 (1H, q, $J=7$); 5.34 (1H, s); 5.46 (1H, t,
 $J=10$); 7.04–7.80 (7H, m); 8.24 (1H, d, $J=8$)



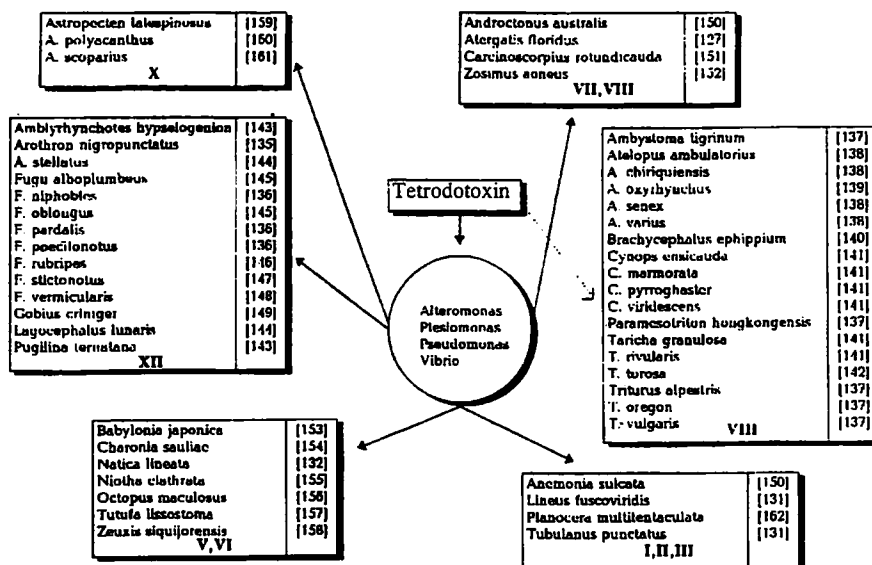
HPLC: [120]
Pharm: anticonvulsive activity

Aspergillus clavatus [125]
C₂₆H₂₄N₄O₆: 488.1696
mp.: 202–204°
[α]_D²⁵: -254° (chl.f.)
UV: 234, 292, 320 (34950, 9550, 6300)
IR (chl.f.): 3525, 1790, 1735, 1715, 1680
Mass: 488 (M⁺)
PMR: 1.24 (3H, d, J=7); 1.31 (3H, d, J=7); 1.59 (3H, d, J=7); 3.09 (1H, dd, J=10,14); 3.47 (1H, dd, J=11,14); 4.12 (1H, quintet, J=7); 4.36 (1H, q, J=7); 5.24 (1H, s); 5.51 (1H, t, J=10); 7.12–7.94 (7H, m); 8.28 (1H, d, J=8)

V. Tetrodotoxins

A distinguishing feature of the distribution of alkaloids of the tetrodotoxin group is their wide representation in marine animals of the most diverse classes and also in amphibia inhabiting bodies of fresh water. In particular, tetrodotoxin and its analogs are found in nemertines (Nemertini: *Lineus*, *Tubulanus*), gastropod mollusks (Gastropoda: *Babylonia*, *Charonia*, *Natica*, *Niotha*, *Tutufa*, *Zeuxis*), a cephalopod mollusk — the blue spotted octopus (Cephalopoda: *Octopus maculosus*), marine crustacea (Crustacea: *Atergatis*, *Zosimus*), globe fishes (Tetraodontidae: *Amblyrhynchotes*, *Arothron*, *Fugu*, *Lagocephalus*, *Pugilina*), amphibian families of salamandrian animals (Ambystomatidae: *Ambystoma*) and true salamanders (Salamandridae: *Cynops*, *Paramesotriton*, *Taricha*, *Triturus*).

Above all, tetrodotoxin is found in corals (Anthozoa: *Anemonia*), turbellarian worms (Turbellaria: *Planocera*), Merostomata (*Androctonus*, *Carcinoscorpius*), starfish (Asteroidea: *Astropecten*), gobies (Gobiidae: *Gobius criniger*), and amphibians of toad families (Brachycephalidae: *Atelopus*, *Brachycephalus*) (see Scheme 1).



Scheme 1. Distribution of tetrodotoxin in Nature: I) Anthozoa (corals); II) Turbellaria; III) Nemertini; V) Gastropoda; VI) Cephalopoda; VII) Crustacea; VIII) Merostomata; X) Asteroidea (starfish); XII) Pisces (fishes); XIII) Amphibia.

According to modern ideas, which have been confirmed experimentally, tetrodotoxin and some of its derivatives are produced by a number of bacteria living on the sea bottom (*Pseudomonas*, *Alteromonas*, *Bacillus*, *Plesiomonas*, *Vibrio*) and are then accumulated by the above-mentioned salt-water dwellers [163, 164]. It is interesting to note that no alkaloids of the

tetrodotoxin group have been detected hitherto either in algae or in higher plants. At the present time, 13 toxins of this type have been described.

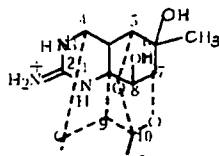
Tetrodotoxin was one of the first quinazoline alkaloids. It was isolated in 1909 from the fish *Fugu rubripes*. However, only after 40 years was tetrodotoxin obtained in crystalline form, while its chemical structure was established by the 1960's and was then confirmed by synthesis [165, 166]. From the chemical point of view, tetrodotoxin is a combination of an aminoperhydroquinazoline with a guanidine group, and it can exist in two forms: hemilactal and lactone [126].

As a rule, these compounds contain hydroxy groups in positions 6 and 8, and sometimes in positions 4 and 9, as well. Other substituents at carbon atom 6 may be hydroxyl, hydrogen, dihydroxymethyl, and methyl. Anhydro bases are known among the tetrodotoxins, and also compounds with extremely original structure, such as 1-hydroxy-5,11-dideoxytetrodotoxin (hydroxyl at a nitrogen atom), tetrodonic acid, and chiriquitoxin.

The group of animal toxins under consideration is known, above all, thanks to their unique biological properties. In very low concentrations (10^{-7} M) tetrodotoxin and its analogs block the transmission of nervous impulses in excitable tissues (nerve and muscle tissues). A large number of original and review papers have been devoted to this question. In spite of the fact that tetrodotoxin possesses a very high toxicity and, from this index, may be assigned to the group of so-called ultratoxins [167], the organisms elaborating them are, as a rule, extremely resistant to them. In view of this, the role of tetrodotoxin and its analogs as powerful agents of chemical protection seems obvious.

As investigations of recent years have shown, tetrodotoxin is present in considerable concentrations in sea-bed deposits [168]. As in many marine animals, the accumulation of this toxin in bottom deposits is the result of the activity of a whole series of tetrodotoxin-producing bacteria — in particular, *Vibrio alginolyticus* [133, 163, 169]. It is assumed that this toxin is a stable component of these deposits and is capable of remaining in an active state for a long time. There is no doubt that tetrodotoxin and its derivatives must exert a definite physiological action on benthic animals feeding on detritus, some of which are apparently capable of accumulating the poison. Thus, the wide distribution of toxins of this type in marine organisms of the most diverse classes has a secondary character.

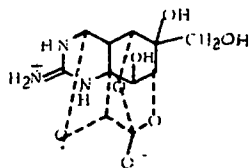
112. 4,9-ANHYDRO-11-DEOXYTETRODOTOXIN



Cynops ensicauda [126]
 $C_{11}H_{15}N_3O_6$: 285.2536
 Amorph.
 Mass (SIMS): 286 (M+H)⁺

PMR ($CD_3COOD+D_2O$ TMS): 1.61 (s, H-11); 2.95 (d, J=3.0, H-4a); 4.01 (t, J=2.0, 2.0, H-7); 4.15 (dd, J=3.0, 2.0, H-5); 4.56 (s, H-9); 4.63 (d, J=2.0, H-8); 5.51 (s, H-4)

113. 4,9-ANHYDROTETRODOTOXIN



Atergatis floridus [127], *Charonia saulnae* [128]
Cynops ensicauda [126], *Fugu parcais*,
F. poecilnotus, *F. rubripes* [129],
F. vermicularis [130], *Lineus fuscovindis* [131],
Natica lineata [132], *Octopus maculosus* [133],
Tubulanus punctatus [131]
 $C_{11}H_{15}N_3O_7$: 301.2526

Amorph.

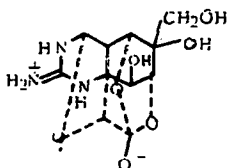
Mass (glycerol): 315.302 (M+H)⁺, 277 (100) [129]

PMR ($CD_3COOD-D_2O$, TMS): 3.06 (d, J=4.1, H-4a); 4.03 (d, J=13.3, 11-CH₂); 4.11 (d, J=13.3, 11-CH₂); 4.27 (q, J=3.5, 1.1, H-7); 4.47 (q, J=4.1, 1.1, H-5); 4.70 (s, H-9); 4.74 (d, J=3.5, H-8); 5.64 (s, H-4)

HPLC: [147]

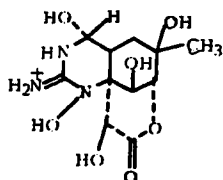
Pharm: LD₅₀-92 Mu/mg (s/c, mice) [129]

114. 4,9-ANHYDRO-6-EPITETRODOTOXIN



Cynops ensicauda [126]
 $C_{11}H_{15}N_3O_7$; 301.2526
 $[\alpha]_D^{+10}$ (AcOH)
 Mass: 302 (M+H)⁻
 PMR: 2.87 (d, J=2.3, H-4a); 3.65, 3.67 (d, J=12.7, 11-CH₂); 4.20 (t, J=2.0, H-7); 4.33 (dd, J=2.3, 2.0, H-5); 4.41 (d, J=2.0, H-8); 4.61 (s, H-9); 5.54 (s, H-4)

115. 1-HYDROXY-5,11-DIDEOXYTETRODOTOXIN

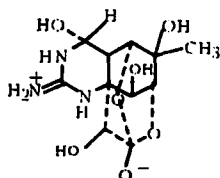


Taricha granulosa [134]
 $C_{11}H_{18}N_3O_7$; 304.2763
 Amorph
 $[\alpha]_D^{+14.47}$ (AcOH)
 Mass: 304.1141 (M⁺)
 PMR: 1.35 (s, H-11); 1.38 (dd, J=15.7, 13.3, H-5β); 2.14 (ddd, J=15.6, 3.7, 1.4, H-5α); 2.77 (ddd, J=13.2, 9.3, 3.9, H-4a); 4.46 (dd, J=3.9, 1.4, H-7); 4.58 (d, J=3.9, H-8); 4.80 (s, H-9); 5.14 (d, J=9.4, H-4)
¹³C NMR:

C-2	158.9	C-6	69.8	C-9	69.9
4	77.5	7	81.2	10	176.6
4a	38.8	8	67.5	11	27.9
5	34.3	8a	74.1		

HPLC

116. 11-DEOXYTETRODOTOXIN



Arothron nigropunctatus [135], *Cynops ensicauda* [126], *C. pyrrhogaster*, *Taricha granulosa*, *Triturus alpestris*, *T. oregon* [137], *Feugu niphobles*, *F. pardalis*, *F. poecilonotus* [136]
 $C_{11}H_{17}N_3O_7$; 303.2684
 mp.: 202° (AcOH-water, dec.)
 $[\alpha]_D^{+5}$ (AcOH)

Mass: 304 (M-H)⁺
 PMR (CD₃COOD+D₂O, TMS): 1.64 (s, H-11); 2.37 (d, J=9.4, H-4a); 3.91 (t, J=1.6, H-7); 3.94 (s, H-9); 4.08 (br.s H-5); 4.30 (d, J=1.6, H-8); 5.49 (d, J=9.4, H-4)

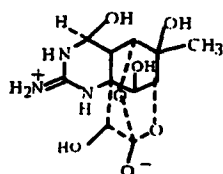
¹³C NMR (CD₃COOD): [126]

C-2	156.4	C-6	69.1	C-9	70.8
4	75.0	7	83.6	10	110.6
4a	40.5	8	72.6	11	25.1
5	77.5	8a	59.1		

HPLC: [209]

Pharm: LD₅₀ 71 mg/kg (i/p, mice)

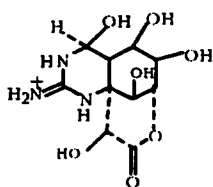
117. 11-DEOXY-4-EPITETRODOTOXIN



Cynops ensicauda [126]
 $C_{11}H_{17}N_3O_7$; 303.2684
 Amorph.
 Mass (SIMS): 304 (M+H)⁻

PMR: 1.64 (s, H-11); 2.87 (d, J=5.0, H-4a); 3.91 (t, J=1.5, 1.5, H-7); 3.96 (s, H-9); 4.13 (d, J=1.5, H-5); 4.29 (d, J=1.5, H-8); 5.13 (d, J=4.9, H-4)

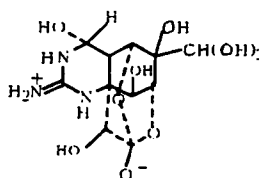
118. 11-NORTETRODOTOXIN-6(R)-OL



Arothron nigropunctatus [135], *Fugu niphobles*, *F. pardalis*, *F. poecilonotus* [136]
 $C_{10}H_{16}N_3O_7$: 290.2495
 Amorph.
 Mass.: 290 (M^-)

PMR ($CD_3COOD-D_2O$, TMS): 2.00 (d, $J=9.8$, H-4a); 3.94 (br.s H-6); 4.15 (d, $J=1.4$, H-8); 4.32 (br.s H-5); 4.56 (s, H-9); 4.72 (br.s H-7); 5.55 (d, $J=9.3$, H-4)
 HPLC.
 Pharm {Ac.}: LD_{99} -70 mg/kg

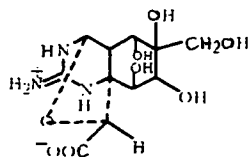
119. 11-OXOTETRODOTOXIN



Arothron nigropunctatus [135]
 $C_{11}H_{17}N_3O_9$: 335.2664
 Amorph.
 Mass.: 336 ($M+H$)⁺

PMR ($CD_3COOD-D_2O$, TMS): 2.31 (br.d $J=9.5$, H-4a); 3.98 (H-9); 4.19 (H-7); 4.27 (H-8); 4.37 (H-5); 5.51 (d, $J=9.5$, H-4); 5.74 (s, H-11)
 HPLC. [209]
 Pharm {Ac.}: LD_{50} 120 mg/kg (i/p, mice)

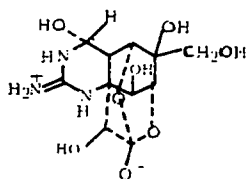
120. TETRODONIC ACID



Feugu pardalis, *F. poecilonotus*, *F. rubripes* [129]
 $C_{11}H_{17}N_3O_8$: 319.2674
 Amorph.
 Mass (glycerol): 320 ($M+H$)⁺, 299, 277 (100)

PMR ($CD_3COOD-D_2O$, TMS): 2.86 (d, $J=4.7$, H-4a); 3.79 (s, H-CH₂); 3.90 (q, $J=4.1$, 1.1, H-7); 4.03 (q, $J=4.1$, 1.1, H-5); 4.29 (d, $J=4.1$, H-8); 5.30 (s, H-9); 5.39 (s, H-4)
 HPLC

121. TETRODOTOXIN



See scheme No. 1
 $C_{11}H_{17}N_3O_8$: 319.2674
 mp: 220°
 Mass (glycerol): 320 ($M+H$)⁺, 100], 302, 277 [129]

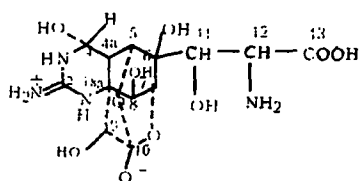
PMR ($CD_3COOD-D_2O$, TMS): 2.35 (d, $J=9.5$, H-4a); 3.96 (s, H-9); 4.02 (d, $J=12.6$, H-11); 4.04 (d, $J=12.6$, H-11); 4.08 (t, $J=1.8$, H-7); 4.25 (br.s, H-5); 5.50 (d, $J=9.4$, H-4)
 ^{13}C NMR (CD_3COOD): [126]

C-2	156.6	C-6	71.5	C-9	70.9
4	75.1	7	79.7	10	110.8
4a	40.7	8	72.8	11	65.5
5	73.8	8a	59.7		

HPLC: [147]

Pharm.: neurotropic activity, vasodilator action. A series of drugs based on tetrodotoxin has been proposed for infiltration and cerebrospinal anesthetics [210].
 LD_{50} 332, 10 mg/kg (oral, i/v mice) [219]

122. CHIRIQUITOXIN



Atelopus chiriquiensis [138]

$C_{13}H_{20}N_4O_{10}$: 392.3178

Amorph.

$[\alpha]_D^{20} -17^\circ$ (AcOH)

IR: 1800, 1741, 1667 [211]

Mass: 393 (M+H)⁺

PMR (CD₃COOD-D₂O, TMS): 2.31 (d, J=9.4, H-4a); 4.00 (s, H-9); 4.27 (d, J=1.8, H-12); 4.39 (br.t H-7); 4.40 (br.s H-5); 4.40 (br.s H-8); 4.90 (d, J=1.8, H-11); 5.51 (d, J=9.4, H-4) [211]

¹³C NMR (CD₃COOD): [138]

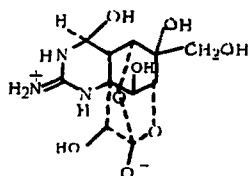
C-2	156.6	C-7	81.1	C-10	111.0
4	75.2	8	72.7	11	70.3
4a	40.5	8a	59.3	12	58.1
5	73.5	9	70.9	13	174.1
6	72.1				

Abs. conf. 11R, 12S [211]

HPLC: [209]

Pharm: {Ac.}: LD₅₀ 14 mg/kg (i/p, mice). Blocks conduction of sodium channels [212]

123. 4-EPITETRODOTOXIN



Arothron nigropunctatus [135], *Charonia sauliae* [128], *Cynops ensicauda* [126], *Fugu pardalis*, *F.poecilnotus*, *F.rubripes* [129]; *Lineus fuscoviridis* [131], *Octopus maculosus* [133]

$C_{11}H_{17}N_3O_8$: 319.2674

Amorph.

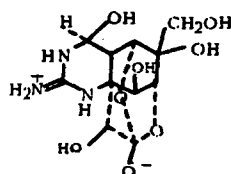
Mass (glycerol): 412, 369, 320 (M+H)⁺, 302, 277 (100)

PMR (CD₃COOD+D₂O, TMS): 2.73 (d, J=5.3, H-4a); 3.85 (vc, H-5); 3.91 (s, 11-CH₂); 3.98 (br.s H-7); 4.07 (br.s H-8); 4.17 (br.s H-9); 5.04 (d, J=5.3, H-4) [129]

HPLC: [147]

Pharm: LD₅₀ 710 μg/mg (s/c, mice) [129]

124. 6-EPITETRODOTOXIN



Ambystoma tigrinum, *Cynops pyrrhogaster*, *Triturus alpestris*, - *T.vulgaris* [137], *Arothron nigropunctatus* [135], *Cynops ensicauda* [126], *Fugu niphobles*, *F.pardalis*, *F.poecilnotus* [136]

$C_{11}H_{17}N_3O_8$: 319.2674

Amorph.

$[\alpha]_D^{20} -5^\circ$ (AcOH)

Mass: 320 (M+H)⁺

PMR (CD₃COOD+D₂O, TMS): 2.01 (d, J=9.0, H-4a); 3.74 (s, H-11); 4.00 (s, H-9); 4.08 (br.s H-7); 4.17 (br.s H-8); 4.30 (d, J=1.6, H-5); 5.55 (d, J=9.4, H-4)

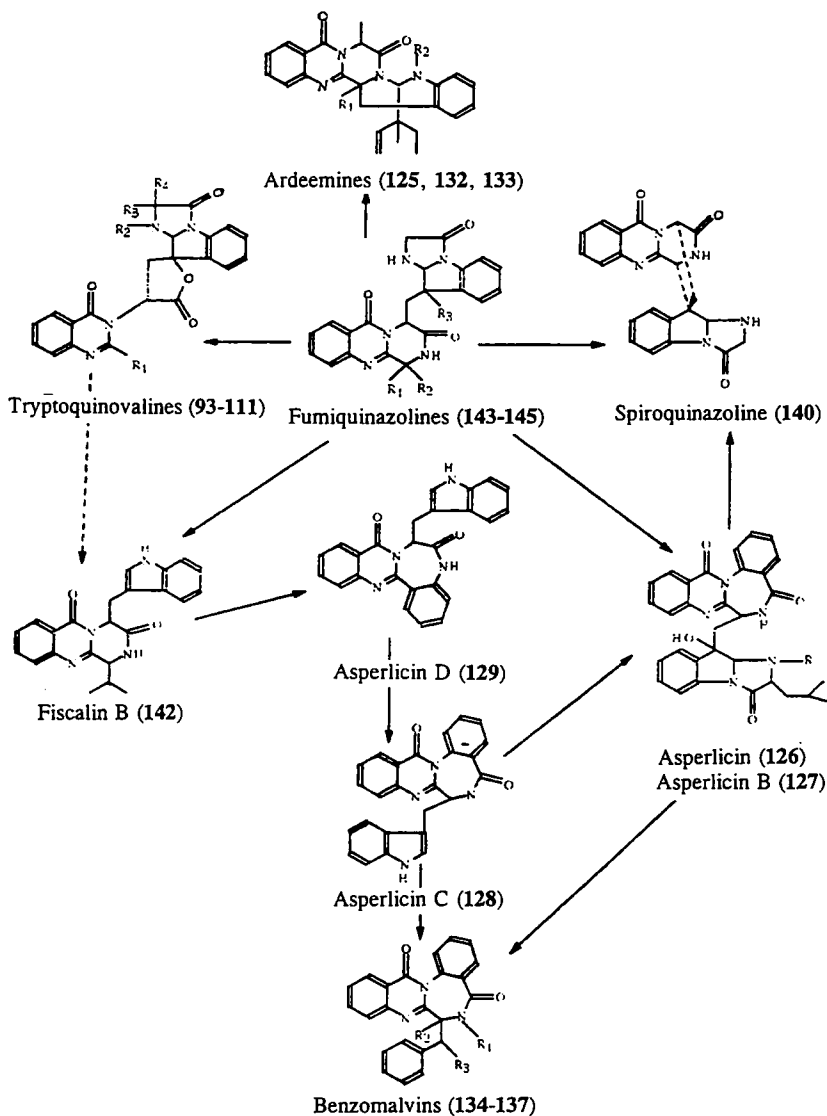
¹³C NMR: CD₃COOD:

C-2	156.5	C-6	72.8	C-9	70.8
4	75.1	7	82.0	10	110.7
4a	41.8	8	72.9	11	65.1
5	75.4	8a	59.6		

Pharm: LD₅₀ 60 mg/kg (i/p, mice)

VI. Quinazoline Alkaloids with Various Structures

In this group we have placed alkaloids differing both in chemical structure and in sources of isolation. Their producing agents are bacteria (*Candida*), ascomycetous fungi (*Apergillus*, *Penicillium*, *Corynascus*, *Neosartorya*), higher plants of the families Polygonaceae (*Polygonum*), Brassicaceae (*Isatis*), Lecythidaceae (*Couroupita*), Acanthaceae (*Strobilanthes*), and marine animals (Bryozoa: *Hincksinoflustra*). So far, 21 such alkaloids are known, and we have arbitrarily combined them into one group.



Scheme 2. Quinazoline alkaloids of ascomycetous fungus (Ascomycetes)

In spite of the obvious structural differences between the alkaloids under consideration, it appears completely possible to distinguish among them a group of compounds closely related both in chemical structure and in natural origin. These are the asperlicins (5 alkaloids), the fumiquinazolines (3 alkaloids), the benzomalvins (4 alkaloids), and the ardeemines (3 alkaloids). The latter have been obtained synthetically quite recently [225]. Fiscalin B and spiroquinazoline must be added to this series. They are all products of the vital activity of the fungi Ascomycetes. In addition to their common origin, the substances mentioned have similar structural elements. Thus, for example, the fumiquinazolines, which represent a combination of pyrazinoquinazoline and imidazoindole heterocyclic systems, are extremely similar to the tryptoquinazolines described in the preceding section. In contrast to the latter, they contain no γ -lactone ring.

The simultaneous presence of fumiquinazolines and tryptoquivalines in *A. fumigatus* probably gives grounds for assuming their biogenetic relationship. On the other hand, in their pyrazinoquinazoline moiety, the fumiquinazolines very

greatly resemble the ardeemines, similarity being observed not only in the structure of the heterocyclic skeleton but also in the positions of the carbonyl groups. Furthermore, in the ardeemines the quinazoline constituent is attached by a C-N bond to a pyrroloindole system. Finally, the fumiquinazolines have an imidazoindole component in common with asperlicin and asperlicin B. However, in the latter it is linked by a methylene bridge with a quinazolinobenzodiazepine fragment. In their turn, with respect to this characteristic, asperlicin and asperlicin B are similar to asperlicins C and D, on the one hand, and to the benzomalvins, on the other hand. Their structural difference consists in the fact that in position 19 asperlicins C and D have an indole grouping and the benzomalvins an unsubstituted aromatic ring.

A recently isolated alkaloid with an original structure — spiroquinazoline — may be considered as a combination of the pyrazinoquinazoline components of the fumiquinazolines and the imidazoindole components of the asperlicins. All these structural features suggest that mutual transitions exist between the tryptoquivalines, fumiquinazolines, benzomalvins, ardeemines, and asperlicins (see Scheme 2). These can quite possibly take place under the conditions of the biocatalytic processes occurring in ascomycetous fungi. This hypothesis is supported by the simultaneous presence of tryptoquivalines and fumiquinazolines in *A. fumigatus* (as mentioned above) and also of tryptoquivalines and fiscalin B in *C. setosus*.

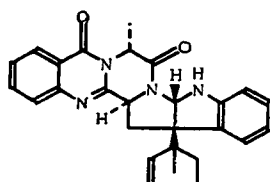
Unfortunately, in view of the clear deficiency of experimental facts, the proposed scheme must be regarded as extremely provisional. It is believed that a further deeper and all-sided study of the biochemistry of the Ascomycetes and, in particular, the genus *Aspergillus* will permit conclusions to be drawn on the biogenesis of this group of alkaloids.

The following two alkaloids — kingdaione (candidine) and tryptanthrine (couroupitine A) are produced both by bacteria and by flowering plants (*Isatis*, *Strobilanthes*, *Couroupita*, *Polygonum*). Both of them are antimicrobial agents [184].

Finally, the last quinazolines with an original structure known at the present time are auranthine from the culture *Penicillium aurantiogriseum* and hinckdentine A from the marine organism *Hincksinoflustra denticulata*. Hinckdentine A is the second representative of the quinazoline alkaloids containing a bromine atom in its molecule.

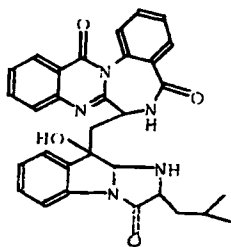
Since practically all quinazoline bases of the group under consideration were isolated comparatively recently (with the exception of tryptanthrin, discovered in 1971), information on their biosynthesis is either completely lacking or has a fragmentary nature. For example, so far as concerns asperlicins C and E, the opinion has been expressed that they originate from tryptophan and anthranilic acid [185]. The recently described synthesis of hinckdentine A from indolo[1,2-c]quinazoline permits the assumption of a similar mechanism for their biosynthesis [186].

125. (-)-ARDEEMINE



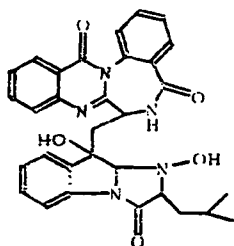
Aspergillus fischeri [213]
Pharm.: [214]

126. ASPERLICIN



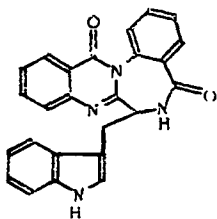
Aspergillus alliaceus [170]
 $C_{31}H_{29}N_5O_4$: 535.5986
XSA:
Pharm: antagonist of the peptide hormone cholecystokinin [171]

127. ASPERLICIN B



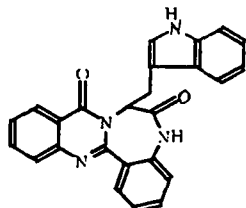
Aspergillus alliaceus [171]
 $C_{31}H_{29}N_5O_5$: 551.5986
Abs. conf.: 2S, 9R*
Pharm: antagonist of cholecystokinin (cck), selectively affecting peripheral cck receptors.

128. ASPERLICIN C



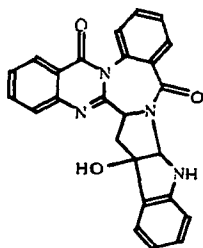
Aspergillus alliaceus [172]
C₂₅H₁₇N₄O₂: 405.4341
XSA: [185]
Pharm: antagonist of the peptide hormone cholecystokinin [171]

129. ASPERLICIN D



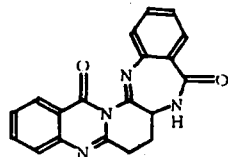
Aspergillus alliaceus [172]
C₂₅H₁₈N₄O₂: 406.4420

130. ASPERLICIN E



Aspergillus alliaceus [172]
C₂₅H₁₈N₄O₃: 422.4410
XSA: [185]
Pharm: antagonist of the peptide hormone cholecystokinin

131. AURANTHINE



Penicillium aurantiogriseum [174]
C₁₉H₁₄N₄O₂: 330.3444
[α]_D²⁰ -164°
{ h-cl. >300° }
UV: 228, 268, 280, 310, 322
IR: 3450, 3175, 1675, 1615, 1595, 1450, 1380, 1250

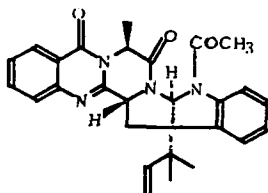
Mass: 330 (M⁺), 277, 249 (100), 248, 234, 220, 192, 132, 130, 124, 119, 102

¹³C NMR:

169.7	130.6	126.9
160.7	129.9	121.1
153.6	129.0	118.6
145.4	128.7	53.2
134.1	127.1	25.4
133.6	127.06	13.6
131.3		

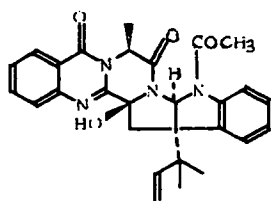
HPLC.

132. (-)-5-N-ACETYLARDEEMINE



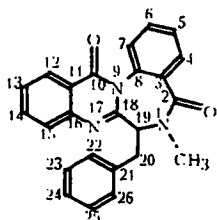
Aspergillus fischeri [213]
C₂₈H₂₅N₄O₃: 465.5293
XSA
Pharm: agent for suppressing the resistance of cancer cells to drugs [225]

133. (-)-15 β -HYDROXY-5-N-ACETYLARDEEMINE



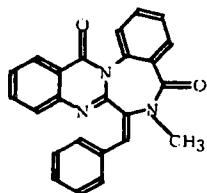
Aspergillus fischeri [213]
C₂₈H₂₈N₄O₄: 484.5520

134. BENZOMALVIN A



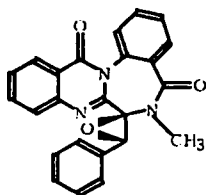
Penicillium sp. [182]
C₂₄H₁₉N₃O₂: 381.4322
PMR: 3.03 (H-27); 3.42, 3.79 (H-20); 4.87 (H-19) [183]
¹³C NMR: 167.6 (C-2); 153.7 (C-18); 59.3 (C-19); 33.7 (C-20); 27.9 (C-27) [183]
HPLC
Pharm: P-substance inhibitor

135. BENZOMALVIN B



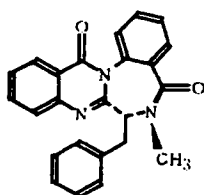
Penicillium sp. [182]
C₂₄H₁₇N₃O₂: 379.4164
PMR: 3.45 (H-27); 6.72 (H-20) [183]
¹³C NMR: 165.8 (C-2); 151.1 (C-18); 128.9 (C-19); 132.8 (C-20); 36.1 (C-27) [183]
HPLC
Pharm: P-substance inhibitor

136. BENZOMALVIN C



Penicillium sp. [182]
C₂₄H₁₇N₃O₃: 395.4154
HPLC
Pharm: P-substance inhibitor

137. BENZOMALVIN D



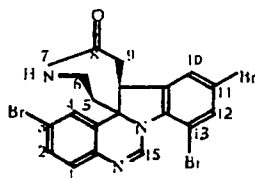
Penicillium sp.: [183]
[α]_D²⁵: +48°
PMR: 2.33 (1H, dd, J=13.8, 11.3); 2.74 (1H, dd, J=13.8, 5.9); 2.91 (3H, s); 4.75 (1H, dd, J=11.3, 5.9); 6.96 (2H, dd, J=7.6, 1.5); 7.20-7.28 (3H, m); 7.53 (1H, td, J=7.6, 1.5); 7.59-7.67 (4H, m); 7.79 (1H, td, J=7.6, 1.5); 8.01 (1H, dd, J=7.6, 1.5); 8.32 (1H, dd, J=7.6, 1.5)

¹³C NMR:

35.9 (t)	127.7 (d)	133.0 (s)
38.3 (q)	128.1 (d)	135.0 (d)
70.3 (d)	128.8 (d, 3C)	135.9 (s)
121.4 (s)	129.0 (d)	146.3 (s)
127.3 (d)	130.7 (d)	153.7 (s)
127.4 (d)	131.1 (d)	161.5 (s)
127.5 (d)	132.1 (s)	165.4 (s)

Abs. conf.: 19S
HPLC

138. HINCKDENTINE A

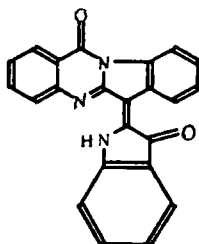


Hincksinoflustra denticulata [175]
 $C_{19}H_{14}Br_3N_3O$: 540.0507
 mp.: 196–197°
 $[\alpha]_D^{25}$ –274°
 UV: 239, 303, 337, 344, 357
 IR: 3400, 1671, 1600, 1576, 1449, 1337, 1300
 Mass: 537, 539, 541, 543 (M^+)

PMR: 1.87 (ddd, J=14.9, 8.8, 1.5, H-5a); 2.17 (ddd, J=14.9, 8.8, 1.5, H-5b); 2.98 (dddd, J=14.9, 8.8, 4.4, 1.5, H-6a); 3.15 (dddd, J=14.9, 5.0, 1.5, 1.5, H-6b); 3.32 (ddd, J=16.1, 6.2, 1.6, H-9a); 3.48 (dd, J=16.1, 3.0, H-9b); 4.07 (dddd, J=6.2, 3.0, 1.4, 1.1, H-9a); 6.58 (br.t 4.4, 1.6, NH); 7.23 (d, J=8.5, H-1); 7.42 (d, J=2.2, H-4); 7.46 (dd, J=1.9, 1.4, H-10); 7.48 (dd, J=8.5, 2.2, H-2); 7.59 (dd, J=1.9, 1.1, H-12); 9.09 (s, H-15)

XSA
 HPLC

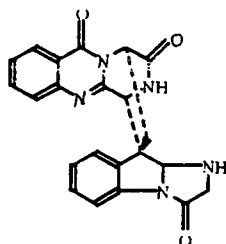
139. KINGDAINONE



Candida lipolytica [176], Isatis tinctoria.
 Strobilanthes cusia [177]
 $C_{23}H_{13}N_3O_2$: 363.3738
 mp.: 269–270°
 UV (chl.f.): 244, 250, 282, 538, 573 (4.34, 4.36, 4.21, 3.95, 4.11) [215]
 IR: 3210, 2730, 1690, 1650, 1625, 1600, 1460, 1355, 1315, 740

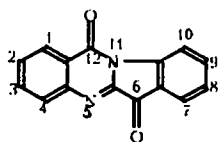
Mass: 364 (23), 363 (100), 335 (37), 259 (10), 258 (11), 242 (9)
 PMR: 7.05 (1H, t, J=7.5, H-5'); 7.09 (1H, d, J=7.8, H-7'); 7.39 (1H, td, J=7.5, 1.2, H-8); 7.48 (1H, td, J=8.0, 1.4, H-9); 7.52 (1H, t, J=7.5, H-2); 7.54 (1H, d, J=8.0, H-6'); 7.75 (1H, dd, J=7.5, 1.0, H-4'); 7.79 (1H, t, J=7.0, H-3); 7.83 (1H, d, J=7.5, H-4); 8.42 (1H, dd, J=7.9, 1.4, H-1); 8.66 (1H, d, J=7.4, H-10); 9.22 (1H, d, J=7.8, H-7); 11.75 (1H, s, NH) [216]

140. SPIROQUINAZOLINE



Aspergillus flavipes [217]

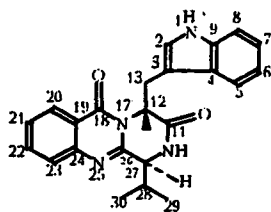
141. TRYPTANTHRIN



Candida lipolytica [178], Couroupita guianensis [179],
 Isatis tinctoria, Polygonum tinctorium [180],
 Strobilanthes cusia [181]
 $C_{15}H_8N_2O_2$: 248.2396
 mp.: 267–268°

UV: 248, 252, 277, 311, 328, 387 (3.36, 3.72, 0.595, 0.729, 0.670, 0.550)
 IR (chl.f.): 1722, 1684, 1596 [181]
 Mass: 248 (M^+ , 100), 220 (32), 192 (13.7)
 PMR: 7.30–8.70 (8H, H-Ar)
 XSA: [220]
 Pharm: fungicidal activity [181]

142. FISCALIN B (CS-D)



Corynascus setosus [208], *Neosartorya fischeri* [218]

$C_{23}H_{22}N_4O_2$: 386.4516

mp: 164.5–170.5° (chlf. meth.)

$[\alpha]_D^{25} -124^\circ$ (meth.)

UV (meth.): 220, 273, 290, 305, 318 (4.65, 4.06, 3.90, 3.60, 3.47)

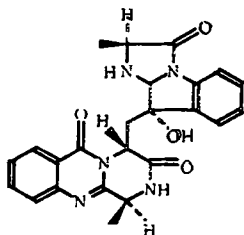
IR: 3258, 1680, 1600, 1460

PMR: 0.57 (3H, d, $J=7.0$, H-29); 0.59 (3H, d, $J=7.0$, H-30); 2.57 (m, H-28); 2.67 (d, $J=2.4$, H-27); 3.56 (dd, $J=14.8$, 5.5, H-13); 3.65 (dd, $J=14.8$, 2.8, H-13); 5.59 (dd, $J=5.5$, 2.8, H-12); 6.10 (br.s, H-10); 6.53 (d, $J=2.1$, H-2); 6.84 (td, $J=7.8$, 1.0, H-6); 7.04 (td, $J=7.8$, 1.0, H-7); 7.21 (br.d, $J=7.8$, H-8); 7.35 (br.d, $J=7.8$, H-5); 7.45 (td, $J=7.9$, 1.1, H-21); 7.48 (br.d, $J=7.9$, H-23); 7.69 (td, $J=7.9$, 1.5, H-22); 8.21 (br.s, H-1); 8.29 (dd, $J=7.9$, 1.5, H-20)

^{13}C NMR:

C-2	122.6 (d)	C-11	168.5 (s)	C-23	126.1 (d)
3	108.2 (s)	12	55.7 (d)	24	146.0 (s)
4	126.2 (s)	13	26.3 (t)	26	149.2 (s)
5	117.6 (d)	18	159.9 (s)	27	57.1 (d)
6	118.9 (d)	19	119.1 (s)	28	28.5 (d)
7	121.5 (d)	20	125.9 (d)	29	13.7 (q)
8	110.0 (d)	21	126.0 (d)	30	17.7 (q)
9	135.0 (s)	22	133.6 (d)		

143. FUMIQUINAZOLINE A

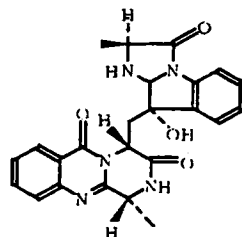


Aspergillus fumigatus [173]

$C_{24}H_{23}N_5O_4$: 445.4752

Pharm: cytotoxic activity

144. FUMIQUINAZOLINE B

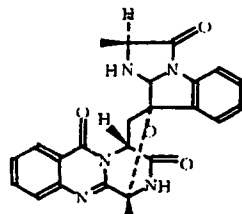


Aspergillus fumigatus [173]

$C_{24}H_{23}N_5O_4$: 445.4752

Pharm: cytotoxic activity

145. FUMIQUINAZOLINE C



Aspergillus fumigatus [173]

$C_{24}H_{21}N_5O_4$: 443.4594

XSA

Pharm: cytotoxic activity

If we consider the problem of the distribution of the quinazoline alkaloids in Nature as a whole, attention is immediately attracted by the circumstance that compounds of this class are found in representatives of all three kingdoms of living Nature — microorganisms, plants, and animals. At the same time, their sources are frequently taxons that are phylogenetically very remote and in no way connected with one another. This suggests that at least some of the quinazoline

alkaloids, especially those of the simplest structure, play the role of protoalkaloids — i.e., substances directly connected with amino acids and found in the most diverse families of plants and animals [90].

At the same time, it is impossible to deny the fact that many quinazolines and some groups of them (for example, the quinazolinocarbolines and tryptoquivalines) are distributed within a single family, genus, or even species. This undoubtedly indicates that they are narrowly specialized and highly organized products of secondary metabolism.

From the aspect of structural chemistry, it appears interesting to us that although the quinazoline alkaloids are a fairly small class of natural compounds, their structures are very diverse. This is shown not only in the methods of constructing the main heterocyclic skeleton but also in the nature of the substituting functional groups, among which are sometimes present such substituents rarely found among alkaloids as bromine, free carboxyl, and others.

In conclusion, it would be desirable to mention that we have scarcely touched upon aspects of structural investigations, stereochemistry, chemical transformations, methods of analysis, pharmacological activity, and many others. Such an urgent problem as the biogenesis of the various types of quinazoline alkaloids has hardly been considered, either. A whole series of reviews and original papers have been devoted to this most important question.

The continuously increasing stream of publications on this subject permits the hope that even in the foreseeable future an answer must be found to the general philosophical question of the place and role in living Nature of alkaloids in general and of the quinazoline alkaloids in particular.

REFERENCES

1. A. Bischler and M. Lang, *Chem. Ber.*, **28**, 279 (1985).
2. W. L. F. Amarego, *Adv. Heterocycl. Chem.*, **24**, 1 (1979).
3. S. Johne and D. Groger, *Pharmazie*, **25**, 22 (1970).
4. C. K. Atal, *Chemistry and Pharmacology of Vasicine — A New Oxytocic and Abortifacient*, Regional Research Laboratory, Jammu-Tawi, New Delhi (1980).
5. S. Johne, in: *The Alkaloids*, Academic Press, New York, Vol. 29, A. Brossi (ed.) (1986), p. 99.
6. J. Bergman, in: *The Alkaloids*, Academic Press, New York, Vol. 21, A. Brossi (ed.) (1983), p. 29.
7. H. S. Mosher, *Ann. N. Y. Acad. Sci.*, **479**, 32 (1986); *Chem. Abstr.*, **106**, 150745 (1987).
8. I. Hermecz and L. Vasvari-Debreczy, *Adv. Heterocycl. Chem.*, **39**, 281 (1986).
9. M. V. Telezhenetskaya, in: *Results of the Investigation of Alkaloid-Bearing Plants [in Russian]*, Kh. N. Aripov (ed.), Fan, Tashkent (1993), p. 221.
10. M. V. Telezhenetskaya and S. Yu. Yunusov, *Khim. Prir. Soedin.*, 731 (1977).
11. M. F. Grundon, *Nat. Prod. Rep.*, **7**, No. 2, 131 (1990).
12. M. F. Grundon, *Nat. Prod. Rep.*, **5**, No. 3, 293 (1988).
13. M. F. Grundon, *Nat. Prod. Rep.*, **4**, No. 3, 225 (1987).
14. L. A. Zenkevich, *The Life of Animals [in Russian]*, Prosveshchenie, Moscow, Vol. 1 (1968).
15. N. A. Krasil'nikov and A. A. Uranov, *The Life of Plants [in Russian]*, Prosveshchenie, Moscow, Vol. 1 (1974).
16. R. N. Chakravati and S. C. Chakravati, *Proc. Inst. Chem.*, **24**, 96 (1952).
17. A. Chatterjee and S. G. Majumdar, *J. Am. Chem. Soc.*, **75**, 4365 (1953).
18. N. Ruangrunsi, P. Tantivatana, R. P. Borris, and G. A. Cordell, *J. Sci. Soc. Thailand*, **7**, 123 (1981).
19. S. C. Pakrashi and J. Bhattacharyya, *J. Sci. Ind Res.*, **21B**, 49 (1962).
20. Y. C. Meinwald, J. Meinwald, and T. Eisner, *Science*, **154**, 390 (1966).
21. H. Schildknecht, W. F. Wenneis, K. H. Weis, and U. Maschwitz, *Z. Naturforsch.*, **21B**, 121 (1966).
22. D. J. Chadwick and I. W. Eaton, *Acta Crystallogr.*, **C**, **39**, 454 (1983).
23. Y. Tezuka, Qung Huang, T. Kikuchi, A. Nishi, and K. Tubaki, *Chem. Pharm. Bull.*, **42**, 2612 (1994).
24. Y. Kimura, T. Inoue, and S. Tamura, *Agric. Biol. Chem.*, **37**, 2213 (1973).
25. M. M. Blight and J. F. Grove, *J. Chem. Soc., Perkin Trans. I*, 1691 (1974).
26. P. J. Suter and W. B. Turner, *J. Chem. Soc., C*, 2240 (1967).
27. K. Prabir Chaudhuri, *J. Nat. Prod.*, **55**, 249 (1992).
28. A. Chatterjee and S. G. Majumdar, *J. Am. Chem. Soc.*, **76**, 2459 (1954).
29. M. Sarkar and D. P. Chakraborty, *Phytochemistry*, **16**, 2007 (1977).

30. T. C. Chou, F. Y. Fu, and Y. S. Kao, *J. Am. Chem. Soc.*, **70**, 1765 (1948).
31. H. Hikino, S. Nabetani, and T. Takemoto, *J. Pharm. Soc. Jpn.*, **93**, 619 (1973).
32. P. K. Chowdhuri, *Phytochemistry*, **26**, 587 (1987).
33. H. Niwa, Y. Yoshida, and K. Yamada, *J. Nat. Prod.*, **51**, 343 (1988).
34. D. L. Dreyer and R. C. Brenner, *Phytochemistry*, **19**, 935 (1980).
35. M. Sarkar and D. P. Chakraborty, *Phytochemistry*, **18**, 694 (1979).
36. S. Mann, *Arch. Mikrobiol.*, **56**, 324 (1967).
37. C. S. Jang, F. Y. Fu, C. Y. Wang, K. C. Huang, G. Lu, and T. C. Chow, *Science*, **103**, 59, (1946).
38. J. B. Koepfli, J. F. Mead, and J. A. Brockman, *J. Am. Chem. Soc.*, **69**, 1837 (1947).
39. F. Ablondi, S. Gordon, J. Morton, and J. H. Williams, *J. Org. Chem.*, **17**, 14 (1952).
40. H. Niwa, M. Watanabe, A. Sano, and K. Yamada, *Tetrahedron*, **50**, 6805 (1994).
41. B. Chowdhury and P. Battacharyya, *Chem. Ind. (London)*, No. 1, 35 (1987).
42. M. P. Jain and T. N. Srivastava, *Fitoterapia*, **57**, 297 (1986).
43. E. Spath and F. Keszler-Gandini, *Monatsh. Chem.*, **91**, 1160 (1960).
44. A. Prakash, R. K. Varma, and S. Ghosal, *Planta Med.*, **43**, 384 (1981).
45. S. Ghosal, P. S. Chauhan, and R. Mehta, *Phytochemistry*, **14**, 830 (1975).
46. M. P. Jain and V. K. Sharma, *Planta Med.*, **46**, 250 (1982).
47. A. H. Amin and D. R. Mehta, *Nature (London)*, **184**, 1317 (1959).
48. D. Kurbanov and B. Kh. Kharekeev, *Khim. Prir. Soedin.*, 685 (1974).
49. V. P. Linichev and A. I. Ban'kovskii, *Tr. Vses. NII Lek. i Arom. Rast.*, 65 (1959).
50. B. Sener and F. Ergun, *Gazi Univ. Eczacilik Fak. Derg.*, **5**, 33 (1988).
51. N. V. Plekhanova and G. P. Sheveleva, *Dokl. Akad. Nauk Kirg.*, 54 (1965).
52. I. Laakso, P. Virkajarvi, H. Airaksinen, and E. Varis, *J. Chromatogr.*, **505**, 424 (1990).
53. Z. Osmanov, A. A. Ibragimov, and S. Yu. Yunusov, *Khim. Prir. Soedin.*, 126 (1982).
54. N. Ya. Koretskaya, *Zh. Obshch. Khim.*, **27**, 3361 (1967).
55. D. Batsurén, M. V. Telezhenetskaya, S. Yu. Yunusov, and T. Ballan, *Khim. Prir. Soedin.*, 418 (1978).
56. T. S. Tulyaganov, *Khim. Prir. Soedin.*, 39 (1993).
57. Kh. N. Khashimov, Author's Abstract of Dissertation ... Candidate of Chemical Sciences [in Russian], Tashkent (1973).
58. Rajbakshmi Poi and N. Adityachaudhury, *J. Indian Chem. Soc.*, **65B**, 814 (1988).
59. V. Yu. Khuzhaev, S. F. Aripova, and R. Sh. Shakirov, *Khim. Prir. Soedin.*, 687 (1994).
60. N. K. Hart, S. R. Johns, and J. A. Lamberton, *Aust. J. Chem.*, **24**, 223 (1971).
61. B. Kh. Zharekeev, M. V. Telezhenetskaya, and S. Yu. Yunusov, *Khim. Prir. Soedin.*, 279 (1973).
62. Kh. N. Khashimov, M. V. Telezhenetskaya, and S. Yu. Yunusov, *Khim. Prir. Soedin.*, 599 (1969).
63. D. Batsurén, M. V. Telezhenetskaya, and S. Yu. Yunusov, *Khim. Prir. Soedin.*, 736 (1980).
64. B. Kh. Zharekeev, Kh. N. Khashimov, M. V. Telezhenetskaya, and S. Yu. Yunusov, *Khim. Prir. Soedin.*, 264 (1974).
65. B. K. Chowdhury and P. Battacharyya, *Phytochemistry*, **24**, 3080 (1985).
66. E. Spath and F. Keszler, *Ber. Dtsch. Chem. Ges.*, **69**, 384 (1936).
67. A. D. Rosenfeld, *Thesen der Ber. zym. VI. Mendeleev-Kongreß. II. Teil, Ausg. V, Verlag Koks und Chemie, Kharkov (1932)*, p. 316.
68. D. Hooper, *Pharm. J.*, **18**, 848 (1881).
69. R. R. Arndt, S. H. Eggers, and A. Jordaan, *Tetrahedron*, **23**, 3521 (1967).
70. H. Kohler, *Biol. Zbl.*, **88**, 165 (1969).
71. S. Johne and D. Groger, *Pharmazie*, **23**, 35 (1968).
72. D. Groger and S. Johne, *Planta Med.*, **13**, 182 (1965).
73. S. Yu. Yunusov and Z. F. Ismailov, *Dokl. Akad. Nauk Uz.*, No. 11, 25 (1956).
74. G. P. Men'shikov, A. I. Ban'kovskii, and V. I. Frolova, *Zh. Obshch. Khim.*, **29**, 3846 (1959).
75. Kh. N. Khashimov, M. V. Telezhenetskaya, Ya. V. Rahkes, and S. Yu. Yunusov, *Khim. Prir. Soedin.*, 453 (1970).
76. K. Schreiber, O. Aurich, and K. Pufahl, *Arch. Pharm.*, **295**, 271 (1962).
77. Kh. N. Khashimov, M. V. Telezhenetskaya, and S. Yu. Yunusov, *Khim. Prir. Soedin.*, 849 (1971).

78. T. S. Tulyaganov, R. Sh. Atadzhanov, N. D. Abdullaev, É. L. Kristallovich, and Z. Osmanov, *Khim. Prir. Soedin.*, **580** (1993).
79. T. S. Tulyaganov, *Khim. Prir. Soedin.*, **87** (1993).
80. T. S. Tulyaganov, *Khim. Prir. Soedin.*, **780** (1994).
81. K. L. Dhar, M. P. Jain, S. K. Koul, and C. K. Atal, *Phytochemistry*, **20**, 319 (1981).
82. M. V. Telezhenetskaya, B. Tashkhodzhaev, M. R. Yagudaev, B. T. Ibragimov, and S. Yu. Yunusov, *Khim. Prir. Soedin.*, **18** (1989).
83. S. Johnne, D. Groger, and M. Hesse, *Helv. Chem. Acta*, **54**, 826 (1971).
84. B. S. Joshi, Y. Bai, M. S. Puar, K. K. Dubose, and S. W. Pelletier, *J. Nat. Prod.*, **57**, 953 (1994).
85. S. R. Johns and J. A. Lambertson, *J. Chem. Soc., Chem. Commun.*, 267 (1965).
86. S. R. Johns, J. A. Lambertson, and H. Soares, *Aust. J. Chem.*, **38**, 1007 (1985).
87. E. S. Pallares, *Arch. Biochem.*, **10**, 235 (1946).
88. K. Pandita, M. S. Bhatia, R. K. Thappa, S. G. Agarwal, K. L. Dhar, and C. K. Atal, *Planta Med.*, **48**, 81 (1983).
89. N. L. Li and J. J. Willaman, *Econ. Botany*, **22**, 239 (1968).
90. G. V. Lazur'evskii and I. V. Terent'eva, *Alkaloids and Plants* [in Russian], Shtiintsa, Kishinev (1975), p. 117.
91. P. P. Waterman, *Biochem. Syst. Ecol.*, **3**, 149 (1975).
92. B. Danieli, G. Palmisano, G. Rainaldi, and G. Russo, *Phytochemistry*, **13**, 1603 (1974).
93. J. F. Ayafor, B. L. Sondengam, and B. T. Ngadjui, *Phytochemistry*, **21**, 2733 (1982).
94. Kwok Ming, A. Gray, P. Waterman, P. But, and Yun-Cheung Kong, *J. Nat. Prod.*, **50**, 1160 (1987).
95. Lin Lie Chwen, Chou Cheng Jen, Kuo Tung, and Chen Chieh Fu, *Zhonghua Yaoxue Zazhi*, **45**, 421 (1993).
96. Y. Asahini and K. Kashiwaki, *J. Pharm. Soc. Jpn.*, **405**, 1293 (1915).
97. I. J. Pachter, R. F. Raffauf, G. E. Ulliyot, and O. Ribeiro, *J. Am. Chem. Soc.*, **82**, 5187 (1960).
98. D. de Correa, O. R. Gottlieb, and A. P. de Padua, *Phytochemistry*, **14**, 2059 (1975).
99. H. Jacobs, F. Ramadaya, S. McLean, M. Perpich-Dumont, F. Puzzuoli, and W. F. Reynolds, *J. Nat. Prod.*, **50**, 507 (1987).
100. H. Banerjee, S. Pal, and N. Adityachaudhury, *Plant Med.*, **55**, 403 (1989).
101. Aim-On Somanabandhu, Nijisiri Ruangrunsi, G. L. Lange, and M. G. Organ, *J. Sci. Soc. Thailand*, **18**, 181 (1992).
102. J. E. T. Corrie, G. H. Green, E. Ritchie, and W. C. Taylor, *Aust. J. Chem.*, **23**, 133 (1970).
103. I. J. Pachter, R. F. Raffauf, G. E. Ulliyot, and O. Ribeiro, *Angew. Chem.*, **69**, 687 (1957).
104. D. de Correa, O. R. Gottlieb, A. P. de Padua, and A. I. da Rocha, *Rev. Latinoamer. Quim.*, **7**, 43 (1976).
105. L. Canonica, B. Danieli, P. Manitto, G. Russo, G. Ferrari, and S. P. A. Simes, *Tetrahedron Lett.*, 4865 (1968).
106. B. Danieli, P. Manitto, F. Ronchetti, G. Russo, and G. Ferrari, *Phytochemistry*, **11**, 1833 (1972).
107. B. Danieli, B. Palmisano, G. Russo, and G. Ferrari, *Phytochemistry*, **12**, 2521 (1973).
108. B. Danieli, C. Farachi, and G. Palmisano, *Phytochemistry*, **15**, 1095 (1976).
109. T. Nakasato, S. Asada, and K. Marui, *J. Pharm. Soc. Jpn.*, **82**, 619 (1962).
110. I. J. Pachter, R. J. Mohrbacher, and D. E. Zacharias, *J. Am. Chem. Soc.*, **83**, 635 (1961).
111. T. Kamikado, S. Murakoshi, and S. Tamura, *Agric. Biol. Chem.*, **42**, 1515 (1978).
112. B. Danieli, G. Lesma, and G. Palmisano, *Experientia*, **35**, 156 (1979).
113. A. Chatterjee, S. Bose, and C. Ghosh, *Tetrahedron*, **7**, 257 (1959).
114. F. Fish, I. A. Meshal, and P. G. Waterman, *Planta Med.*, **29**, 310 (1976).
115. N. Shoji, A. Umeyama, A. Iuchi, N. Saito, T. Takemoto, K. Nomoto, and Y. Ohizumi, *J. Nat. Prod.*, **51**, 791 (1988).
116. B. S. Joshi, M. S. Puar, K. M. Moore, and S. W. Pelletier, *Heterocycles*, 1365 (1991).
117. M. F. Faskhutdinov, M. V. Telezhenetskaya, and N. D. Abdullaev, unpublished.
118. N. Shoji, A. Umeyama, A. Iuchi, N. Saito, T. Takemoto, K. Nomoto, and Y. Ohizumi, *J. Nat. Prod.*, **51**, 161 (1988).
119. B. Danieli, P. Manitto, F. Ronchetti, G. Russo, and G. Ferrari, *Experientia*, **28**, 249 (1972).
120. G. Buchi, Kin Chun Luk, B. Kobbe, and J. M. Townsend, *J. Org. Chem.*, **42**, 244 (1977).
121. M. Yamazaki, H. Fujimoto, and E. Okuyana, *Chem. Pharm. Bull.*, **25**, 2554 (1977).
122. M. Yamazaki, H. Fujimoto, and E. Okuyana, *Chem. Pharm. Bull.*, **26**, 111 (1978).
123. M. Yamazaki, E. Okuyama, and Y. Maebayashi, *Chem. Pharm. Bull.*, **27**, 1611 (1979).

124. J. P. Springer, *Tetrahedron Lett.*, 339 (1979).
125. J. Clardy, J. P. Springer, G. Buchi, K. Matsuo, and R. Wightman, *J. Am. Chem. Soc.*, **97**, 663 (1975).
126. T. Yasumoto, M. Yotsu, and M. Murata, *J. Am. Chem. Soc.*, **110**, 2344 (1986).
127. T. Noguchi, J. K. Jeon, O. Arakawa, H. Sugita, Y. Deguchi, Y. Shida, and K. Hashimoto, *J. Biochem. (Tokyo)*, **99**, 331 (1986).
128. H. Narita, H. Masumoto, S. Mizuno, T. Noguchi, T. Saito, Y. Shida, and K. Hashimoto, *Shokuhin Eiseigaku Zasshi*, **30**, 522 (1989).
129. M. Nakamura and T. Yasumoto, *Toxicon*, **23**, 271 (1985).
130. T. Noguchi, D. F. Hwang, O. Arakawa, H. Sugita, Y. Deguchi, Y. Shida, and K. Hashimoto, *Mar. Biol.*, **94**, 625 (1987).
131. K. Miyazawa, M. Higashiyama, K. Ito, T. Noguchi, O. Arakawa, Y. Shida, and K. Hashimoto, *Toxicon*, **26**, 867 (1988).
132. Deng Fwu Hwang, Chuang Hsun Chueh, and Sen Shyong Jeng, *Toxicon*, **28**, 21 (1990).
133. D. F. Hwang, O. Arakawa, T. Saito, T. Noguchi, U. Shimidu, K. Tsukamoto, Y. Shida, and K. Hashimoto, *Mar. Biol.*, **100**, 327 (1989).
134. K. Yuichians and S. Yuzuru, *J. Am. Chem. Soc.*, **115**, 827 (1993).
135. S. Khora and T. Yasumoto, *Tetrahedron Lett.*, 4393 (1989).
136. A. Endo, S. Khora, M. Murata, H. Naoki, and T. Yasumoto, *Tetrahedron Lett.*, 427 (1988).
137. M. Yotsu, M. Iorizzi, and T. Yasumoto, *Toxicon*, **28**, 238 (1990).
138. Y. H. Kim, G. B. Brown, H. S. Mosher, and F. A. Fuhrman, *Science*, **189**, 151 (1975).
139. D. Mebs and K. Schmidt, *Toxicon*, **27**, 819 (1989).
140. A. Sebben, C. A. Schwartz, D. Valente, and E. G. Mendes, *Toxicon*, **24**, 799 (1986).
141. M. S. Brown and H. S. Mosher, *Science*, **140**, 295 (1963).
142. V. C. Twitty and H. H. Johnson, *Science*, **80**, 78 (1934).
143. Y. Fuchi, S. Morisaki, T. Nagata, K. Simazaki, N. Ohtomo, T. Noguchi, and K. Hashimoto, *Oita-ken Kogai Eisei Nenpo*, **15**, 35 (1987).
144. M. Toyoda, A. Kingkate, C. Jaengsawang, W. Thanissorn, and C. Halilamian, *J. Food Saf.*, **3**, 207 (1981).
145. D. F. Hwang, T. Noguchi, O. Arakawa, T. Abe, and K. Hashimoto, *Nippon Suisan Gakkaishi*, **54**, 2001 (1988).
146. Y. Tawara, *J. Pharm. Soc. Jpn.*, **29**, 587 (1909).
147. T. Yasumoto and T. Michishita, *Agric. Biol. Chem.*, **49**, 3077 (1985).
148. M. Arisawa, T. Horiuchi, T. Hayashi, Y. Tezuka, T. Kikuchi, and N. Morita, *Chem. Pharm. Bull.*, **41**, 1472 (1993).
149. Y. Hashimoto and T. Noguchi, *Toxicon*, **9**, 79 (1971).
150. G. H. De Vries and M. Lazdunski, *J. Biol. Chem.*, **257**, 11684 (1982).
151. A. Kingsuwan, Y. Nagashima, T. Noguchi, Y. Shida, S. Suvapeepan, P. Suwansakornkul, and K. Hashimoto, *Bull. Jpn. Soc. Sci. Fish.*, **53**, 261 (1987).
152. D. Yasumura, Y. Oshima, T. Yasumoto, A. C. Alcalá, and L. C. Alcalá, *Agric. Biol. Chem.*, **50**, 593 (1986).
153. T. Noguchi, J. Maruyama, Y. Ueda, K. Hashimoto, and T. Harata, *Nippon Suisan Gakkaishi*, **47**, 909 (1981).
154. H. Narita, T. Noguchi, J. Maruyama, Y. Ueda, K. Hashimoto, Y. Watanabe, and K. Hida, *Nippon Suisan Gakkaishi*, **47**, 935 (1981).
155. J. K. Jeon, H. Narita, M. Nara, T. Noguchi, J. Maruyama, and K. Hashimoto, *Nippon Suisan Gakkaishi*, **50**, 2099 (1984).
156. D. Scheumack, M. E. H. Howden, I. Spence, and R. J. Quinn, *Science*, **199**, 188 (1978).
157. T. Noguchi, J. Maruyama, Y. Ueda, H. Narita, and K. Hashimoto, *Toxicon*, **22**, 219 (1984).
158. H. Narita, T. Noguchi, J. Maruyama, M. Nara, and K. Hashimoto, *Nippon Suisan Gakkaishi*, **50**, 85 (1984).
159. J. Maruyama, T. Noguchi, J. K. Jeon, T. Harada, and K. Hashimoto, **40**, 1395 (1984).
160. T. Noguchi, H. Narita, J. Maruyama, and K. Hashimoto, *Bull. Jpn. Soc. Sci. Fish.*, **48**, 1173 (1982).
161. K. Miyazawa, T. Noguchi, J. Maruyama, J. K. Jeon, M. Otsuka, and K. Hashimoto, *Mar. Biol.*, **90**, 61 (1985).
162. K. Miyazawa, J. K. Jeon, J. Maruyama, T. Noguchi, K. Ito, and K. Hashimoto, *Toxicon*, **24**, 645 (1968).
163. E. V. Thuesen and K. Kogure, *Biol. Bull.*, **176**, 191 (1989).
164. M. Yotsu, T. Yamazaki, Y. Meguro, A. Endo, M. Murata, H. Nacki, and T. Yasumoto, *Toxicon*, **25**, 225 (1987).

165. T. Goto, Y. Kishi, S. Takahashi, and Y. Hirata, *Tetrahedron*, **21**, 2059 (1965).
166. R. B. Woodward, *Pure Appl. Chem.*, **9**, 49 (1964).
167. B. N. Orelov and D. B. Gelashvili, *Zootoxinology (Poisonous Animals and Their Venoms)* [in Russian], Vysshaya Shkola, Moscow (1985).
168. K. Kogure, H. K. Do, and E. V. Thuesen, *Mar. Ecol. Prod. Ser.*, **45**, 303 (1988).
169. T. Matsui, S. Taketsugu, K. Kodama, A. Ishii, K. Yamamori, and C. Shimizu, *Nippon Suisan Gakkaishi*, **55**, 2199 (1989).
170. M. A. Goetz, M. Lopez, R. L. Monaghan, R. S. L. Chang, V. J. Lotti, and T. B. Chen, *J. Antibiot.*, **38**, 1633 (1985).
171. R. S. L. Chang, V. J. Lotti, R. L. Monaghan, J. Birnbaum, E. O. Stapley, M. A. Goetz, G. Albers-Schonberg, A. A. Patchett, J. M. Liesch, O. D. Hensens, and J. P. Springer, *Science*, **230**, 177 (1985).
172. M. A. Goetz, R. L. Monaghan, R. S. L. Chang, J. Ondeyka, T. B. Chen, and V. J. Lotti, *J. Antibiot.*, **41**, 875 (1988).
173. A. Numata, C. Takahashi, T. Matsushita, T. Miyamoto, K. Kawai, Y. Usami, E. Matsumura, M. Inoue, H. Ohishi, and T. Shingu, *Tetrahedron Lett.*, 1621 (1991).
174. S. E. Yeulet, P. G. Mantle, J. N. Bilton, H. S. Rzepa, and R. N. Sheppard, *J. Chem. Soc., Perkin Trans. I*, 1891 (1986).
175. A. J. Blackman, T. W. Hambley, K. Picker, W. C. Taylor, and N. Thirasasana, *Tetrahedron Lett.*, 5561 (1987).
176. H. P. Fiedler, Dissertation, Tübingen (1974).
177. J.-Ch. Zou and L. Huang, *Acta Pharm. Sinica*, **20**, 45 (1985).
178. E. Fiedler, H. P. Fiedler, A. Gerhard, W. K. Schierlein, W. A. König, and H. Zahner, *Arch. Mikrobiol.*, **79**, 187 (1971).
179. J. Bergman, B. Egestad, and J.-O. Lindstrom, *Tetrahedron Lett.*, 2625 (1977).
180. G. Honda, V. Tosirisuk, and M. Tabata, *Planta Med.*, **38**, 275 (1980).
181. G. Honda and M. Tabata, *Planta Med.*, **36**, 85 (1979).
182. H. H. Sun, C. J. Barrow, D. M. Sedlock, A. M. Gillum, and R. Cooper, *J. Antibiot.*, **47**, 515 (1994).
183. H. H. Sun, C. J. Barrow, and R. Cooper, *J. Nat. Prod.*, **58**, 1575 (1995).
184. L. A. Mitscher, Wai-Cheong Wong, T. De Meulenaere, J. Sulko, and S. Drake, *Heterocycles*, **15**, 1017 (1981).
185. M. G. Bock, R. M. Di Pardo, S. M. Pitzemberger, C. F. Homnick, J. P. Springer, and R. M. Freidinger, *J. Org. Chem.*, **52**, 1644 (1987).
186. A. D. Billimoria and M. P. Cava, *J. Org. Chem.*, **59**, 6777 (1994).
187. N. Mohr, H. Budzikiewicz, H. Korth, and G. Pulverer, *Liebigs Ann. Chem.*, **1515**, 187 (1981).
188. S. C. Pakrashi and J. Bhattacharyya, *Tetrahedron*, **24**, 1 (1968).
189. S. C. Pakrashi, J. Bhattacharyya, L. F. Johnson, and H. Budzikiewicz, *Tetrahedron*, **19**, 1011 (1963).
190. T. Kametani, Chu Van Loc, T. Higa, M. Koizumi, M. Ihara, and K. Fukumoto, *J. Am. Chem. Soc.*, **99**, 2306 (1977).
191. T. Kametani, Chu Van Loc, T. Higa, M. Ihara, and K. Fukumoto, *J. Chem. Soc., Perkin Trans. I*, 2347 (1977).
192. H. Schildknecht, U. Maschwitz, and W. F. Wenneis, *Naturwiss.*, **54**, 196 (1967).
193. J. Bergman and A. Brynolf, *Tetrahedron*, **46**, 1295 (1990).
194. L. Li, C. Z. Qiao, X. L. Li, and T. Y. Dong, *Yaoxue Xuebao*, **28**, 238 (1993).
195. M. Kato, M. Inaba, H. Itahana, E. Ohara, and K. Nakamura, *Chem. Abstr.*, **115**, 64012 (1991).
196. C. Kaneko, T. Chiba, K. Kasai, and C. Miwa, *Heterocycles*, **23**, 1385 (1985).
197. H. Culbertson, J. G. Decius, and B. E. Christensen, *J. Am. Chem. Soc.*, **74**, 4834 (1952).
198. Y. Nakagawa and R. V. Stevens, *J. Org. Chem.*, **53**, 1873 (1988).
199. T. Onaka, *Tetrahedron Lett.*, 4387 (1971).
200. J. S. Fitzgerald, S. R. Johns, J. A. Lambertson, and A. H. Redcliffe, *Aust. J. Chem.*, **19**, 151 (1966).
201. J. Bergman and S. Bergman, *J. Org. Chem.*, **50**, 1246 (1985).
202. Y. Kano, M. Yasuda, K. Saito, and K. Komatsu, *Shoyakugaku Zasshi*, **43**, 339 (1989).
203. M. C. M. Yang, S. L. Wu, J. S. Kuo, and C. F. Chen, *European J. Pharm.*, **182**, 537 (1990).
204. C. I. King, Y. C. Kong, N. S. Wong, H. W. Yeung, H. H. S. Fong, and U. Sanakawa, *J. Nat. Prod.*, **51**, 245 (1980).

205. T. Kametani, T. Higa, K. Fukumoto, and M. Koizumi, *Heterocycles*, **4**, 23 (1976).
206. J. Tamas, G. Bujtas, K. Horvath-Dora, and O. Clauder, *Acta Chim. Acad. Sci. Hung.*, **89**, 85 (1976).
207. T. Kametani, T. Higa, Chu Van Loc, M. Ihara, M. Koizumi, and K. Fukumoto, *J. Am. Chem. Soc.*, **98**, 6186 (1976).
208. H. Fujimoto, E. Negishi, K. Yamaguchi, N. Nishi, and M. Yamazaki, *Chem. Pharm. Bull.*, **44**, 1843 (1996).
209. M. Yotsu, A. Endo, and T. Yasumoto, *Agric. Biol. Chem.*, **53**, 893 (1989)
210. I. S. Azhgikhin, V. G. Gandel', S. D. Mekhtikhanov, N. V. Serebryannikov, N. A. Aksenova, and V. V. Finkel', *Farmatsiya*, No. 4, 27 (1979).
211. M. Yotsu, T. Yasumoto, Y. H. Kim, H. Naoki, and C. Y. Kao, *Tetrahedron Lett.*, 3187 (1990).
212. C. Y. Kao, P. N. Yeoh, M. D. Goldfinger, F. A. Fuhrman, and H. S. Mosher, *J. Pharmacol. Exp. Therap.*, **217**, 416 (1981).
213. J. E. Hochlowski, M. M. Mullally, S. G. Spanton, D. N. Whittern, P. Hill, and J. B. McAlpine, *J. Antibiot.*, **46**, 380 (1993).
214. J. P. Karwowski, M. Jackson, R. P. Rasmussen, P. E. Humphrey, J. B. Poddig, W. L. Kohl, M. H. Scherr, S. Kadam, and J. B. McAlpine, *J. Antibiot.*, **46**, 374 (1993).
215. J. Bergman and U. Tilstam, *Tetrahedron*, **41**, 2883 (1985).
216. H. Ladtsch and H. Ludwig-Kohn, *Liebigs Ann. Chem.*, 1847 (1986).
217. C. J. Barrow and H. H. Sun, *J. Nat. Prod.*, **57**, 471 (1994).
218. S. M. Wong, L. L. Musza, G. C. Kydd, R. Kullnig, A. M. Gillum, and R. Cooper, *J. Antibiot.*, **46**, 545 (1993).
219. U. Lindequist and E. Teuscher, *Pharmazie*, **42**, 1 (1987).
220. W. Fedeli and F. Mazza, *J. Chem. Soc., Perkin Trans. II*, 1621 (1974).
221. T. Kametani, Chu Van Loc, M. Ihara, and K. Fukumoto, *Heterocycles*, **9**, 1585 (1978).
222. M. Uskokovic, J. Iacobelli, V. Toome, and W. Wenner, *J. Org. Chem.*, **29**, 582 (1964).
223. Wen-Fei Chiou, Jyh-Fei Liao, and Chieh-Fu Chen, *J. Nat. Prod.*, **59**, 374 (1996).
224. J. Yamahara, T. Yamada, T. Kitani, Y. Naitoh, and H. Fujimura, *Chem. Pharm. Bull.*, **37**, 1820 (1989).
225. S. P. Marsden, K. M. Depew, and S. J. Danishefsky, *J. Am. Chem. Soc.*, **116**, 11143 (1994).
226. B. R. Baker, J. McEvoy, R. E. Schaub, J. P. Joseph, and J. H. Williams, *J. Org. Chem.*, **18**, 178 (1953)
227. R. Shakirov, M. V. Telezhenetskaya, I. A. Bessonova, S. F. Aripova, I. A. Israelov, M. N. Sultankhodzhaev, V. I. Vinogradova, V. I. Akhmedzhanova, T. S. Tulyaganov, B. T. Salimov, and V. A. Tel'nov, *Khim. Prir. Soedin.*, a) 468; b) 990; c) 297; d) 298; e) 298; f) 422; g) 424; h) 425; i) 443; j) 461; k) 763; l) 767; m) 767; n) 798; o) 798; p) 798; r) 799; s) 799 (1996).